

**Credit Distribution for PG Microbiology
First Year
Semester-I**

Part	Course	Course Title	Credit	No. of Hours
Part I	Core I	General Microbiology and Microbial Diversity	5	7
	Core II	Immunology, Immunomics and Microbial Genetics	5	7
	Core III	Practical-I	4	6
	Elective I	Forensic Science/ Health Hygiene/ Microalgal Technology	3	5
	Elective II	Bioinstrumentation/ Herbal Technology and Cosmetic Microbiology / Essentials of Laboratory Management and Biosafety	3	5
Total			20	30

**First Year
Semester-II**

Part	Course	Course Title	Credit	No. of Hours
Part I	Core IV	Medical Bacteriology and Mycology	5	6
	Core V	Medical Virology and Parasitology	5	6
	Core VI	Practical-II	4	6
	Elective III	Epidemiology/ Clinical Diagnostic Microbiology/ Bioremediation	3	4
	Elective IV	Bioinformatics/ Nanobiotechnology/ Clinical Research and Clinical Trials	3	4
Part II	Skill Enhancement Course I	Vermitechnology	2	4
	Compulsory Paper	Human rights	2	2
	MOOC course	Online course	2	-
Total			26	30

**Second Year
Semester-III**

Part	Course	Course Title	Credit	No. of Hours
Part I	Core VII	Soil and Environmental Microbiology	5	6
	Core VIII	Recombinant DNA Technology and Biotechnology	5	6
	Core IX	Practicals III	5	6
	Core X Industry Module	Fermentation Technology and Pharmaceutical Microbiology	4	6
	Elective V	Biosafety, Bioethics and IPR/ Toxinology/ Water Conservation and Water Treatment	3	3
Part II	Skill Enhancement Course II	Organic Farming and Biofertiliser Technology	2	3
		Internship / Industrial Activity	2	-
			26	30

**Second Year
Semester-IV**

Part	Course	Course Title	Credit	No. of Hours
Part I	Core XI	Food & Dairy Microbiology	5	6
	Core XII	Research Methodology & Biostatistics	5	6
	Project	Project with Viva Voce	7	10
	Elective VI	Bioenergy/ Marine Microbiology/ Life Science for Competitive Examinations	3	4
Part II	Skill Enhancement Course	Microbial Quality Control and Testing	2	4
Part III	Extension Activity		1	-
			23	30
Total			95	120

Consolidated Table for Credit Distribution

	Category of Courses	Credits for each courses	Number of Courses	Number of Credits in each Category of courses	Total Credits	Total Credits for the Programme
Part I	Core Theory	5	8	40	82	92(CGPA)
	Core practical	4	3	12		
		5	1	5		
	Project with viva-voce	7	1	7		
	Discipline centric/Generic centric elective	3	6	18		
Part II	Skill Enhancement	2	3	6	10	3(Non CGPA)
	Human rights	2	1	2		
	MOOC course	2	1	2		
	Summer Internship	2	1	2	3	
Part III	Extension activity	1	1	1		

Subject Code	Subject Name	Category	L	T	P	S	Credits	Inst. Hours	Marks		
									CIA	External	Total
22MBP GCT1	General Microbiology and Microbial Diversity	Core Course I	Y	Y	-	-	4	6	25	75	100
Course Objectives											
CO1	Acquire knowledge on the principles of different types of microscopes and their applications.										
CO2	Compare and contrast the structure of bacteria and fungi. Illustrate nutritional requirements and growth in bacteria.										
CO3	Exemplify, isolate and cultivate microalgae from diverse environmental sources.										
CO4	Explain various pure culture techniques and discuss sterilization methods.										
CO5	Discuss the importance and conservation of microbial diversity.										
UNIT	Details								No. of Hours	Course Objectives	
I	History and Scope of Microbiology. Microscopy – Principles and applications. Types of Microscopes - Bright field, Dark-field, Phase-contrast, Fluorescence microscope, Transmission electron microscope (TEM) and Scanning electron microscope (SEM). Sample preparation for SEM & TEM. Atomic force, Confocal microscope. Micrometry – Stage, Ocular and its applications.								20	CO1	
II	Bacterial Structure, properties and biosynthesis of cellular components – Cell wall. Actinomycetes and Fungi - Distribution, morphology, classification, reproduction and economic importance. Sporulation. Growth and nutrition - Nutritional requirements, Growth curve, Kinetics of growth, Batch culture, Synchronous growth, Measurement of growth and factors affecting growth.								20	CO2	
III	Algae - Distribution, morphology, classification, reproduction and economic importance. Isolation of algae from soil and water. Media and methods used for culturing algae, Strain selection and large-scale cultivation. Life cycle - <i>Chlamydomonas</i> , <i>Volvox</i> (Green algae), <i>Nostoc</i> (Cyanobacteria), <i>Sargassum</i> (Brown algae), <i>Polysiphonia</i> , (Red algae).								15	CO3	
IV	Microbial techniques - Sterilization, Disinfection and its validation. Staining methods – Simple, Differential and Special staining. Automated Microbial identification systems - Pure cultures techniques – Cultivation of Anaerobic organisms. Maintenance and preservation of pure cultures. Culture collection centres - National and International.								15	CO4	
V	Biodiversity - Introduction to microbial biodiversity – Classification and applications of Thermophiles , Alkaliphiles								20	CO5	

	Acidophiles , Barophiles and Halophiles. Methanogenes / Conservation of Biodiversity.		
		Total	90
Course Outcomes			
Course Outcomes	On completion of this course, students will;		
CO1	Examine various microbes employing the microscopic techniques learnt. Measure and compare the size of microbes.	PO1, PO4, PO11	
CO2	Differentiate and appreciate the anatomy of various microbes. Plan the growth of microbes for different environmental conditions.	PO1, PO4	
CO3	Identify and cultivate the algae understanding their habitat. Analyze the morphology, classify and propagate depending on its economic importance.	PO7, PO8, PO9	
CO4	Create aseptic conditions by following good laboratory practices.	PO3, PO4, PO7	
CO5	Categorize and cultivate a variety of extremophiles following standard protocols for industrial applications.	PO5, PO7, PO8, PO9	
Text Books			
1.	Kanunga R. (2017). Ananthanarayanan and Panicker's Text book of Microbiology. (10 th Edition). Universities Press (India) Pvt. Ltd.		
2.	Chan E.C.S., Pelczar M. J. Jr. and Krieg N. R. (2010). Microbiology. (5 th Edition). Mc.Graw Hill. Inc, New York.		
3.	Prescott L. M., Harley J. P. and Klein D. A. (2004). Microbiology. (6 th Edition). McGraw - Hill company, New York.		
4.	White D. Drummond J. and Fuqua C. (2011). The Physiology and Biochemistry of Prokaryotes, Oxford University Press, Oxford, New York.		
5.	Dubey R.C. and Maheshwari D. K. (2009). Textbook of Microbiology. S. Chand, Limited.		
REFERENCES BOOKS			
1.	Tortora G. J., Funke B. R. and Case C. L. (2015). Microbiology: An Introduction (12 th Edition). Pearson, London, United Kingdom		
2.	Webster J. and Weber R.W.S. (2007). Introduction to Fungi. (3 rd Edition). Cambridge University Press, Cambridge.		
3.	Schaechter M. and Leaderberg J. (2004). The Desk encyclopedia of Microbiology. Elseiver Academic Press, California.		
4.	Ingraham, J.L. and Ingraham, C.A. (2000) Introduction to Microbiology. (2 nd Edition). Books / Cole Thomson Learning, UK.		
5.	Madigan M. T., Bender K.S., Buckley D. H. Sattley W. M. and Stahl (2018) Brock Biology of Microorganisms. (15 th Edition). Pearson.		
Web Resources			
1.	http://sciencenetlinks.com/tools/microbeworld		
2.	https://www.microbes.info/		
3.	https://www.asmscience.org/VisualLibrary		
4.	https://open.umn.edu/opentextbooks/BookDetail.aspx?bookId=404		
5.	https://www.grsmu.by/files/file/university/cafedry//files/essential_microbiology.pdf		
Methods of Evaluation			
	Continuous Internal Assessment Tests		

Internal Evaluation	Assignments	25 Marks
	Seminars	
	Attendance and Class Participation	
External Evaluation	End Semester Examination	75 Marks
	Total	100 Marks
Methods of Assessment		
Recall (K1)	Simple definitions, MCQ, Recall steps, Concept definitions	
Understand / Comprehend (K2)	MCQ, True/False, Short essays, Concept explanations, Short summary or overview	
Application (K3)	Suggest idea/concept with examples, Suggest formulae, Solve problems, Observe, Explain	
Analyze (K4)	Problem-solving questions, Finish a procedure in many steps, Differentiate between various ideas, Map knowledge	
Evaluate (K5)	Longer essay/ Evaluation essay, Critique or justify with pros and cons	
Create (K6)	Check knowledge in specific or offbeat situations, Discussion, Debating or Presentations	

Mapping with Programme Outcomes

	PO 1	PO 2	PO 3	PO 4	PO 5	PO 6	PO 7	PO 8	PO 9	PO 10	PO 11	PO 12	PO 13	PO 14
CO1	M			M							S			
CO2	L			S										
CO3							S	S	M					
CO4			S	S			S							
CO5					S		S	S	S					

**FIRST YEAR
SEMESTER-I**

Subject Code	Subject Name	Category	L	T	P	S	Credits	Inst. Hours	Marks		
									CI A	External	Total
22MBPGC T2	Immunology, Immunomics and Microbial Genetics	Core Course II	Y	Y	-	-	4	6	25	75	100
Course Objectives											
CO1	Discuss immunity, organs and cells involved in immunity. Compare the types of antigens and their properties.										
CO2	Describe immunoglobulin and its types. Categorize MHC and understand its significance.										
CO3	Elucidate the mechanisms of different hypersensitivity reactions. List out the Vaccines and discuss their development.										
CO4	Acquire knowledge the structure DNA in prokaryotes and eukaryotes										
CO5	Explain out gene transfer studies in microbes.										
UNIT	Details								No. of Hours	Course Objectives	
I	Introduction to biology of the immune system – Cells and organs of Immune System. T and B lymphocytes – Origin, development, differentiation, lymphocyte subpopulation in humans. Innate immunity and Acquired immunity – Active and Passive immunity. Antigens - features associated with antigenicity and immunogenicity. Basis of antigen specificity. MHC genes and products, Structure of MHC molecules. Antigen processing and presentation to T- lymphocytes.								20	CO1	
II	Immunoglobulins. Theories of antibody production. Class switching and generation of antibody diversity. Monoclonal and polyclonal antibodies. Complement system – mode of activation- Classical, Alternate and Lectin pathways, biological functions. Antigen recognition – TCR. Physiology of acquired immune response – various phases of HI, CMI – Cell mediated cytotoxicity, DTH response.								20	CO2	
III	Hypersensitivity – Types and mechanisms, Autoimmunity, Tumor Immunity and Transplantation immunology. Immunodeficiency-Primary immunodeficiency and Secondary immunodeficiencies. Genetics of Immunohematology – Genetic basis and significance of ABO and Rh System. Diagnostic Immunology - Precipitation reaction, Immunodiffusion methods - SRID, ODD. Immunoelectrophoresis - Rocket and Counter current								25	CO3	

	electrophoresis. Agglutination- Labeled Assay- Immunofluorescence assay, Radio immunoassay, ELISA. Role of cytokines, lymphokines and chemokines. Introduction to Vaccines and Adjuvants - Types of vaccines. Immunomics - Introduction and Applications. Antigen engineering for better immunogenicity and use for vaccine development-multiepitope vaccines. Reverse vaccinology.		
IV	Structure of prokaryotic and eukaryotic genome. Introduction to prokaryotic genomic structure, Eukaryotic Genome - Structure of chromatin, chromosome, centromere, telomere, nucleosome. Modifications- methylation, acetylation, phosphorylation.	13	CO4
V	Gene Transfer Mechanisms- Conjugation and its uses. Transduction, Generalized and Specialized, Transformation- Natural Competence and Transformation. Transposition, Mechanism and Types of Transposition reactions.	12	CO5
	Total	60	

Course Outcomes

Course Outcomes	On completion of this course, students will;	
CO1	Categorize the immune response to a variety of antigens. Identify different immune cells involved in immunity.	PO1, PO4, PO6, PO7, PO9
CO2	Justify the significance of MHC molecules in immune response and antibody production.	PO1, PO4, PO5, PO6, PO9
CO3	Design antibodies and evaluate immunological assays in patient samples.	PO4, PO6, PO7, PO8, PO9, PO10
CO4	Analyze genomic DNA of prokaryotes and eukaryotes.	PO4, PO5, PO6, PO7, PO9, PO10
CO5	Summarize gene transfer mechanisms for experimental study.	PO4, PO5, PO6, PO7, PO9, PO10

Text Books

1.	Coico R., Sunshine G. and Benjamini E. (2003). Immunology – A Short Course. (5 th Edition). Wiley-Blackwell, New York.
2.	Owen J. A., Punt J., Stranford S. A. and Kuby J. (2013). Immunology, (7 th Edition). W. H. Freeman and Company, New York.
3.	Abbas A. K., Lichtman A. H. and Pillai S. (2021). Cellular and Molecular Immunology. (10 th Edition). Elsevier.
4.	Malacinski G.M. (2008). Freifelder's Essentials of Molecular Biology. (4 th Edition). Narosa Publishing House, New Delhi.
5.	Gardner E. J. Simmons M. J. and Snusted D.P. (2006). Principles of Genetics. (8 th Edition). Wiley India Pvt. Ltd.

References Books

1.	Travers J. (1997). Immunobiology - The Immune System in Health and Disease. (3 rd Edition). Current Biology Ltd. New York.
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2.	Delves P.J., Martin S., Burton D. R. and Roitt I. M. (2006). Roitt's Essential Immunology. (11 th Edition). Wiley-Blackwell.
3.	Hay F. C. and Westwood O. M. R. (2002). Practical Immunology (4 th Edition). Wiley-Blackwell.
4.	Glick B. R. and Patten C.L. (2018). Molecular Biotechnology – Principles and Applications of Recombinant DNA. (5 th Edition). ASM Press.
5.	Russell P.J. (2010). Genetics - A Molecular Approach. (3 rd Edition). Pearson New International Edition.

Web Resources

1.	https://www.ncbi.nlm.nih.gov/books/NBK279395/
2.	https://med.stanford.edu/immunol/phd-program/ebook.html
3.	https://ocw.mit.edu/courses/hst-176-cellular-and-molecular-immunology-fall-2005/pages/lecture-notes/
4.	[PDF] Lehninger Principles of Biochemistry (8 th Edition) By David L. Nelson and Michael M. Cox Book Free Download - StudyMaterialz.in
5.	https://microbenotes.com/gene-cloning-requirements-principle-steps-applications/

Methods of Evaluation

Internal Evaluation	Continuous Internal Assessment Tests	25 Marks
	Assignments	
	Seminars	
	Attendance and Class Participation	
External Evaluation	End Semester Examination	75 Marks
	Total	100 Marks

Methods of Assessment

Recall (K1)	Simple definitions, MCQ, Recall steps, Concept definitions
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Mapping with Programme Outcomes

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO 10	PO 11	PO 12	PO 13	PO 14
CO1	S			M		M	S		S					
CO2	S			S	M	S			S					
CO3				S		S	S	S	S	M				
CO4				S	M	S	M		S	M				
CO5				S	M	S	M		S	S				

**FIRST YEAR
SEMESTER-I**

Subject Code	Subject Name	Category	L	T	P	S	Credits	Inst. Hours	Marks		
									CIA	External	Total
22MBP GCP1	Practical I	Core Course III- Practical I	-	-	Y	-	4	6	25	75	100
Course Objectives											
CO1	Gain knowledge on the fundamentals, handling and applications of microscopy, sterilization methods. Identify microbes by different staining methods.										
CO2	Prepare media for bacterial growth. Discuss plating and growth measurement techniques.										
CO3	Acquire adequate skills to perform blood grouping and serological reactions.										
CO4	Provide fundamental skills in preparation, separation and purification of immunoglobulin.										
CO5	Apply the knowledge of molecular biology skills in clinical diagnosis.										
UNIT	Details								No. of Hours	Course Objectives	
I	<p>Microscopic Techniques: Light microscopy: Hay infusion broth. Wet mount to show different types of microbes, hanging drop.</p> <p>Dark field microscopy – Motility.</p> <p>Washing and cleaning of glass wares: Sterilization methods: moist heat, dry heat, and filtration.</p> <p>Quality control check for each method.</p> <p>Staining techniques - Simple staining, Gram’s staining, Acid fast staining, Spore, Capsule, Flagella.</p>								20	CO1	
II	<p>Media Preparation: Preparation of liquid, solid and semisolid media. Agar deeps, slants, plates. Preparation of basal, enriched, selective and enrichment media.</p> <p>Preparation of Biochemical test media, media to demonstrate enzymatic activities.</p> <p>Microbial Physiology: Purification and maintenance of microbes. Streak plate, pour plate, and slide culture technique. Aseptic transfer.</p> <p>Direct counts – Total cell count, Turbidometry. Viable count - pour plate, spread plate. Bacterial growth curve. Anaerobic culture methods.</p>								20	CO2	
III	<p>Hematological reactions - Blood Grouping – forward and reverse, Rh Typing</p> <p>Identification of various immune cells by morphology – Leishman staining.</p> <p>Agglutination Reactions- Latex Agglutination reactions- RF, ASO.</p> <p>Detection of HBs Ag by ELISA.</p> <p>Precipitation reactions in gels– Ouchterlony double immunodiffusion (ODD) and Mancini’s single radial immunodiffusion (SRID)</p>								20	CO3	

	Immuno-electrophoresis - Rocket immuno electrophoresis and counter current immuno electrophoresis.		
IV	Preparation of lymphocytes from peripheral blood by density gradient centrifugation.	10	CO4
V	Western Blotting – Demonstration. Isolation of genomic DNA from <i>E. coli</i> and analysis by agarose gel electrophoresis Estimation of DNA using colorimeter (Diphenylamine reagent) Separation of proteins by polyacrylamide gel electrophoresis (SDS-PAGE) UV induced mutation and isolation of mutants by replica plating technique. Plasmid DNA isolation from <i>E.coli</i> . RNA isolation from yeast. RNA estimation by Orcinol method.	20	CO5
	Total	60	
Course Outcomes			
Course Outcomes	On completion of this course, students will;		
CO1	Apply microscopic techniques and staining methods in the identification and differentiation of microbes.	PO1, PO6, PO7, PO8, PO9, PO11	
CO2	Apply the knowledge on the sterilization of glass wares and media by different methods and measurement of cell growth.	PO1, PO6, PO7, PO8, PO9, PO11	
CO3	Perform and evaluate immunological reactions to aid diagnosis.	PO5, PO7, PO8, PO9, PO11	
CO4	Assess the level of lymphocytes in a blood sample and purify immunoglobulin employing appropriate techniques.	PO6, PO7, PO8, PO9, PO11	
CO5	Perform DNA extraction and gene transfer mechanisms, analyze and identify by gel electrophoresis	PO6, PO7, PO8, PO9, PO11	
Text Books			
1.	Dubey R.C. and Maheshwari D. K. (2010). Practical Microbiology. S. Chand.		
2.	Cappuccino, J. and Sherman, N. (2002). Microbiology: A Laboratory Manual, (6 th Edition). Pearson Education, Publication, New Delhi.		
3.	Cullimore D. R. (2010). Practical Atlas for Bacterial Identification. (2 nd Edition). -Taylor & Francis.		
4.	Rich R. R., Fleisher T. A., Shearer W. T., Schroeder H, Frew A. J. and Weyand C. M. (2018). Clinical Immunology: Principles and Practice. (5 th Edition). Elsevier.		
5.	Glick B. R. and Patten C.L. (2018). Molecular Biotechnology – Principles and Applications of Recombinant DNA. (5 th Edition). ASM Press.		
References Books			
1.	Collee J. G., Fraser A.G. Marmion B. P. and Simmons A. (1996). Mackie & McCartney Practical Medical Microbiology. (14 th Edition). Elsevier, New Delhi.		
2.	Gupta P. S. (2003). Clinical Immunology. Oxford University Press.		
3.	Brown T.A. (2016). Gene Cloning and DNA Analysis. (7 th Edition). John Wiley and Jones, Ltd.		

4.	Dale J. W., Schantz M.V. and Plant N. (2012). From Gene to Genomes – Concepts and Applications of DNA Technology. (3 rd Edition). John Wileys and Sons Ltd. 2012.
5.	Maloy S. R., Cronan J.E. Jr. and Freifelder D. (2011). Microbial Genetics. (2 nd Edition). Narosa Publishing Home Pvt Ltd.

Web Resources

1.	http://textbookofbacteriology.net/
2.	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC149666/
3.	https://ocw.mit.edu/courses/hst-176-cellular-and-molecular-immunology-fall-2005/pages/lecture-notes/
4.	[PDF] Lehninger Principles of Biochemistry (8 th Edition) By David L. Nelson and Michael M. Cox Book Free Download - StudyMaterialz.in
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Methods of Evaluation

Internal Evaluation	Continuous Internal Assessment Tests	25 Marks
	Attendance and Class Participation	
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Recall (K1)	Simple definitions, MCQ, Recall steps, Concept definitions
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Mapping with Programme Outcomes

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO 10	PO 11	PO 12	PO 13	PO 14
CO1	M					S	M	M	S		M			
CO2	M					S	M	M	S		M			
CO3					S		S	M	S		M			
CO4						S	S	M	S		S			
CO5						S	S	M	S		S			

Subject Code	Subject Name	Category	L	T	P	S	Credits	Inst. Hours	Marks		
									CIA	External	Total
22MBPGE 1A	Forensic Science	Elective Course I (Choice -1)	3	1	-	-	3	4	25	75	100
Course Objectives											
CO1	Understand the Scope, need and learn the tools and techniques in forensic science.										
CO2	Comprehend organizational setup of a forensic science laboratory.										
CO3	Identify and Examine body fluids for identification.										
CO4	Extract DNA from blood samples for investigation.										
CO5	Recognize medico legal post mortem procedures and their importance.										
UNIT	Details								No. of Hours	Course Objectives	
I	Forensic Science - Definition, history and development of forensic science. Scope and need of forensic science in present scenario. Branches of forensic science. Tools and techniques of forensic science. Duties of a forensic scientist.								12	CO1	
II	Forensic science laboratories - Organizational setup of a forensic science laboratory. Central and State level laboratories in India. Mobile forensic science laboratory and its functions. Forensic microbiology - Types and identification of microbial organisms of forensic significance.								12	CO2	
III	Forensic serology - Definition, identification and examination of body fluids - Blood, semen, saliva, sweat and urine. Forensic examination and identification of hair and fibre.								12	CO3	
IV	DNA profiling - Introduction, history of DNA typing. Extraction of DNA from blood samples - Organic and Inorganic extraction methods. DNA fingerprinting - RFLP, PCR, STR. DNA testing in disputed paternity.								12	CO4	
V	Forensic toxicology - Introduction and concept of forensic toxicology. Medico legal post mortem and their examination. Poisons - Types of poisons and their mode of action.								12	CO5	
	Total								60		
Course Outcomes	On completion of this course, students will;										
CO1	Identify the scope and need of forensic science in the present scenario.								PO1, PO6, PO7, PO8, PO9		
CO2	Plan for the organizational setup and functioning of forensic science laboratories.								PO1, PO6, PO7, PO8, PO9		
CO3	Analyze the biological samples found at the crime scene.								PO1, PO5, PO7, PO8, PO9		
CO4	Perform extraction and identification of DNA obtained from body fluids.								PO1, PO6, PO7, PO8, PO9		
CO5	Discuss the concept of forensic toxicology.								PO1, PO6, PO7, PO8, PO9		
Text Books											

1.	Nanda B. B. and Tewari R. K. (2001) Forensic Science in India: A Vision for the Twenty First Century. Select Publishers, New Delhi. ISBN- 10:8190113526 / ISBN-13:9788190113526.	
2.	James S. H. and Nordby, J. J. (2015) Forensic Science: An Introduction to Scientific and Investigative Techniques. (5 th Edition). CRC Press. ISBN-10:9781439853832 / ISBN-13:978-1439853832.	
3.	Li R. (2015) Forensic Biology. (2 nd Edition). CRC Press, New York. ISBN-13:978-1-4398-8972-5.	
4.	Sharma B.R (2020) Forensic science in criminal investigation and trials. (6 th Edition)Universal Press.	
5.	Richard Saferstein (2017). Criminalistics- An introduction to Forensic Science. (12 th Edition).Pearson Press.	
Reference books		
1.	Nordby J. J. (2000). Dead Reckoning. The Art of Forensic Detection- CRC Press, New York. ISBN:0-8493-8122-3.	
2.	Saferstein R. and Hall A. B. (2020). Forensic Science Hand book, Vol. I, (3 rd Edition). CRC Press, New York. ISBN-10:1498720196.	
3.	Lincoln, P.J. and Thomson, J. (1998). (2 nd Edition). Forensic DNA Profiling Protocols. Vol. 98. Humana Press. ISBN: 978-0-89603-443-3.	
4.	Val McDermid (2014). Forensics. (2 nd Edition). ISBN 9780802125156.	
5.	Vincent J. DiMaio., Dominick DiMaio. (2001). Forensic Pathology (2 nd Edition). CRC Press.	
Web resources		
1.	http://clsjournal.ascls.org/content/25/2/114	
2.	https://www.ncbi.nlm.nih.gov/books/NBK234877/	
3.	https://www.elsevier.com/books/microbial-forensics/budowle/978-0-12-382006-8	
4.	https://www.researchgate.net/publication/289542469_Methods_in_microbial_forensics	
5.	https://cisac.fsi.stanford.edu/events/microbial_forensics	
Methods of Evaluation		
Internal Evaluation	Continuous Internal Assessment Tests	25 Marks
	Assignments	
	Seminars	
	Attendance and Class Participitation	
External Evaluation	End Semester Examination	75 Marks
	Total	100 Marks

Methods of Assessment	
Recall (K1)	Simple definitions, MCQ, Recall steps, Concept definitions
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Analyse (K4)	Problem-solving questions, Finish a procedure in many steps, Differentiate between various ideas, Map knowledge
Evaluate (K5)	Longer essay/ Evaluation essay, Critique or justify with pros and cons
Create (K6)	Check knowledge in specific or offbeat situations, Discussion, Debating or Presentations

Mapping with Programme Outcomes

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO 10	PO 11	PO 12	PO 13	PO 14
CO1	L					S	M	M	S					
CO2	M					S	M	M	S					
CO3	L				S		S	M	S					
CO4	M					S	S	M	S					
CO5	M					S	S	M	S					

Subject Code	Subject Name	Category	L	T	P	S	Credits	Inst. Hours	Marks		
									CIA	External	Total
22MBP GE1B	Health and Hygiene	Elective Course I (Choice- 2)	Y	Y	-	-	3	4	25	75	100
Course Objectives											
CO1	Acquire knowledge on hygiene and live healthy.										
CO2	Provide insights on health laws for food safety and hygiene.										
CO3	Explain health, physical exercises and their importance.										
CO4	Illustrate mental hygiene and involved in mental hygiene.										
CO5	Describe the various health and health education programmes by the government.										
UNIT	Details								No. of Hours	Course Objectives	
I	Introduction to hygiene and healthful live. Factors affecting health, health habits and practices. Recognizing positive & negative practices in the community. Scientific principles related to health.								12	CO1	
II	Nutrition and Health – Balanced diet, Food surveillance, food Fortification, adulteration and preventive measures. Health laws for food safety. Environmental and housing hygiene. Ventilation and lighting.								12	CO2	
III	Physical health, physical exercises and their importance – Walking, jogging, yoga and meditation, stress relief. International control of health, WHO. Personal hygiene, Sun bathing, Colon Hygiene. Health destroying habits and addictions - Pan, supari, ganja, drinking, smoking, tea and coffee.								12	CO3	
IV	Mental hygiene - factors responsible, developmental tasks, basic needs, emotional stability. Mental hygiene and health in infancy, early childhood, adolescence, adulthood and old age. Mental health occupational hazards.								12	CO4	
V	Health programme and health education – Malaria control, Tuberculosis control, AIDS control programmes and Immunization Programmes. Family planning, Reproductive and Child health programmes (RCH).								12	CO5	
	Total								60		
Course Outcomes											
Course Outcomes	On completion of this course, students will;										
CO1	Identify factors affecting health and health habits.								PO1, PO5, PO10		

CO2	Execute the knowledge of ventilation and lighting. Justify Health laws for food safety and hygiene.	PO5, PO10
CO3	Follow personal hygiene to avoid diseases and Prevent people from health-destroying habits and addictions.	PO5, PO10
CO4	Explore Mental hygiene and maintain emotional stability.	PO5, PO10
CO5	Participate in health education programmes	PO1, PO5, PO10
Text Books		
1.	Bamji M. S., Krishnaswamy K. and Brahmam G. N. V. (2019). Textbook of Human Nutrition. (4 th Edition). Oxford and IBH Publishing Co. Pvt. Ltd., New Delhi	
2.	Swaminathan (1995) Food& Nutrition (Vol I) (2 nd Edition). The Bangalore Printing &Publishing Co Ltd., Bangalore.	
3.	Paniker J. C. K. and Ananthanarayan R. (2017). Textbook of Microbiology. (10 th Edition). Universities Press (India) Pvt. Ltd	
4.	Lindsay Dingwall .(2010). Personal Hygiene Care Print ISBN:9781405163071 Online ISBN:9781444318708 DOI:10.1002/9781444318708	
5.	Walter C. C. Pakes(1900). The Science of Hygiene: a Text-book of Laboratory Practice. (London: Methuen and Co.,).	
References Books		
1.	Khader V. (2000) Food, Nutrition and Health, Kalyan Publishers, New Delhi.	
2.	Srilakshmi, B. (2010) Food Science, (5 th Edition) New Age International Ltd., New Delhi.	
3.	Dubey R.C. and Maheshwari D. K. (2010). Practical Microbiology. S. Chand.	
4.	Park K. 2007, Park's text book of Preventive and Social Medicine, Banarsidas Bhanot publishers, India.	
5.	Srilakshmi, 2002, Dietetics, New Age Publications, India	
Web Resources		
1.	Health and Hygiene - Personal Hygiene, Community Hygiene and Diseases (vedantu.com)	
2.	Chapter-32.pdf (nios.ac.in)	
3.	Menstrual Health and Hygiene Guide Student Health and Counseling Services (ucdavis.edu)	
4.	https://nap.nationalacademies.org/read/11756/chapter/13	
5.	http://ecoursesonline.iasri.res.in/mod/page/view.php?id=112325	
Methods of Evaluation		
Internal Evaluation	Continuous Internal Assessment Tests	25 Marks
	Assignments	
	Seminars	
	Attendance and Class Participation	
External Evaluation	End Semester Examination	75 Marks

	Total	100 Marks
Methods of Assessment		
Recall (K1)	Simple definitions, MCQ, Recall steps, Concept definitions	
Understand / Comprehend (K2)	MCQ, True/False, Short essays, Concept explanations, Short summary or overview	
Application (K3)	Suggest idea/concept with examples, Suggest formulae, Solve problems, Observe, Explain	
Analyse (K4)	Problem-solving questions, Finish a procedure in many steps, Differentiate between various ideas, Map knowledge	
Evaluate (K5)	Longer essay/ Evaluation essay, Critique or justify with pros and cons	
Create (K6)	Check knowledge in specific or offbeat situations, Discussion, Debating or Presentations	

Mapping with Programme Outcomes

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO 10	PO 11	PO 12	PO 13	PO 14
CO1	L				S					M				
CO2					S					M				
CO3					S					L				
CO4					S					M				
CO5	L				S					M				

Subject Code	Subject Name	Category	L	T	P	S	Credits	Inst. Hours	Marks		
									CIA	External	Total
22MBP GE1C	Microalgal Technology	Elective Course I (Choice -3)	Y	Y	-	-	3	4	25	75	100
Course Objectives											
CO1	Characterize the different groups of algae.										
CO2	Describe the cultivation and harvesting of algae.										
CO3	Identify the commercial applications of various algal products.										
CO4	Apply microalgae for environmental applications.										
CO5	Employ microalgae as alternate fuels.										
UNIT	Details								No. of Hours	Course Objectives	
I	Introduction to Algae - General characteristics. Classification of algae. Salient features of different groups of algae. Distribution - Freshwater, brackish water and marine algae. Identification methods.								12	CO1	
II	Cultivation of freshwater and marine microalgae - Growth media. Isolation and enumeration of microalgae. Laboratory cultivation and maintenance. Outdoor cultivation - Photobioreactors - construction, types and operation; raceway ponds - Heterotrophic and mixotrophic cultivation - Harvesting of microalgae biomass.								12	CO2	
III	Microalgae in food and nutraceutical applications - Algal single cell proteins. Cultivation of <i>Spirulina</i> . Microalgae as aquatic, poultry and cattle feed. Microalgal biofertilizers. Value-added products from microalgae. Pigments - Production of microalgal carotenoids and their uses. Microalgal secondary metabolites - Pharmaceutical and cosmetic applications.								12	CO3	
IV	Microalgae in environmental applications. Phycoremediation - Domestic and industrial waste water treatment. High-rate algal ponds and surface-immobilized systems - Treatment of gaseous wastes by microalgae. Algal blooms, algicides for algal control.								12	CO4	
V	Microalgae as feed stock for production of biofuels - Carbon-neutral fuels. Lipid-rich algal strains - <i>Botryococcus braunii</i> . Drop-in fuels from algae - hydrocarbons and biodiesel, bioethanol, biomethane, biohydrogen and syngas from microalgae biomass.								12	CO5	
	Total								60		

Course Outcomes		
Course Outcomes	On completion of this course, students will;	
CO1	Acquire knowledge in the field of microalgal technology and their characteristics.	PO1
CO2	Identify the methods of algal cultivation and harvesting.	PO1, PO6
CO3	Recognize and recommend the use of microalgae as food, feed and fodder.	PO7, PO8, PO9
CO4	Promote microalgae in phycoremediation.	PO7, PO9, PO11, PO14
CO5	Compare and critically evaluate recent applied research in these microalgal applications.	PO7, PO8, PO9
Text Books		
1.	Lee R.E. (2008). Phycology. Cambridge University Press.	
2.	Sharma O.P. (2011). Algae. Tata McGraw-Hill Education.	
3.	Shekh A., Schenk P., Sarada R. (2021). Microalgal Biotechnology. Recent Advances, Market Potential and Sustainability. Royal Society of Chemistry.	
4.	Lele. S.S., Jyothi Kishen Kumar (2008). Algal bio process technology. New Age International P(Ltd)	
5.	Das., Mihirkumar. Algal Biotechnology. Daya Publishing House, New Delhi.	
References Books		
1	Andersen R.A. (2005). Algal culturing techniques. Academic Press, Elsevier.	
2	Bux F. (2013). Biotechnological Applications of Microalgae: Biodiesel and Value-added Products. CRC Press.	
3	Singh B., Baudhdh K., Bux, F. (2015). Algae and Environmental Sustainability. Springer.	
4	Das D. (2015). An algal biorefinery: An integrated approach. Springer.	
5	Bux F. and Chisti Y. (2016). Algae Biotechnology: Products and Processes. Springer.	
Web Resources		
1	https://www.classcentral.com/course/algae-10442	
2	https://onlinecourses.nptel.ac.in/noc19_bt16/preview	
3	https://freevidelectures.com/course/4678/nptel-industrial-biotechnology/46	
4	https://nptel.ac.in/courses/103103207	
5.	https://www.sciencedirect.com/topics/earth-and-planetary-sciences/microalgae	
Methods of Evaluation		
Internal Evaluation	Continuous Internal Assessment Tests	
	Assignments	
	Seminars	
	Attendance and Class Participation	
External Evaluation	End Semester Examination	75 Marks
	Total	100 Marks

Methods of Assessment	
Recall (K1)	Simple definitions, MCQ, Recall steps, Concept definitions
Understand / Comprehend (K2)	MCQ, True/False, Short essays, Concept explanations, Short summary or overview
Application (K3)	Suggest idea/concept with examples, Suggest formulae, Solve problems, Observe, Explain
Analyse (K4)	Problem-solving questions, Finish a procedure in many steps, Differentiate between various ideas, Map knowledge
Evaluate (K5)	Longer essay/ Evaluation essay, Critique or justify with pros and cons
Create (K6)	Check knowledge in specific or offbeat situations, Discussion, Debating or Presentations

Mapping with Programme Outcomes

	PO 1	PO 2	PO 3	PO 4	PO 5	PO 6	PO 7	PO 8	PO 9	PO 10	PO 11	PO 12	PO 13	PO 14
CO1	S													
CO2	S					M								
CO3							S	S	S					
CO4							S		S		M			M
CO5							M	S	S					

Subject Code	Subject Name	Category	L	T	P	S	Credits	Inst. Hours	Marks		
									CIA	External	Total
22M BPG E2A	Bioinstrumentation	Elective Course II (Choice -1)	Y	Y	-	-	3	4	25	75	100
Course Objectives											
CO1	Explain the principles and working mechanisms of laboratory instruments.										
CO2	Discuss chromatography techniques and molecular biology techniques.										
CO3	Illustrate molecular techniques in biological applications.										
CO4	Acquire knowledge on spectroscopic techniques										
CO5	Demonstrate the use of radio isotopes in various techniques.										
UNIT	Details								No. of Hours	Course Objectives	
I	Basic laboratory Instruments. Aerobic and anaerobic incubator – Biosafety Cabinets - Fume Hood, pH meter, Lyophilizer, Flow cytometry. Centrifugation techniques: Basic principles of centrifugation - Standard sedimentation coefficient - measurement of sedimentation co-efficient; Applications in determination of molecular weight.								12	CO1	
II	General principles of chromatography - Chromatographic Performance parameters; Types- Thin layer chromatography, Paper Chromatography, Adsorption, ion exchange, Gel filtration, affinity. Flash Chromatography and Ultra Performance convergence chromatography. Two dimensional chromatography. Stimulated moving bed chromatography (SEC).								12	CO2	
III	Electrophoresis: General principles - moving boundary electrophoresis - electrophoretic mobility – supportive materials – electro endosmosis – types (horizontal, vertical and two dimensional electrophoresis) - Principle and applications - paper electrophoresis, starch gel electrophoresis, Disc gel, Agarose gel, SDS – PAGE, Immuno electrophoresis. Blotting techniques -Southern, northern and western blotting.								12	CO3	
IV	Spectroscopic techniques: Principle, simple theory of absorption of light by molecules, electromagnetic spectrum, instrumentation and application of UV- visible, Raman, FTIR spectrophotometer, spectrofluorimetry,								12	CO4	

	Atomic Absorption Spectrophotometer, Flame spectrophotometer, NMR, ESR, Emission Flame Photometry and GC-MS. Detection of molecules in living cells - FISH and GISH..		
V	Radioisotopic techniques: Principle and applications of tracer techniques in biology. Radioactive isotopes - radioactive decay; Detection and measurement of radioactivity. Geiger- Muller and Scintillation counters, auto radiography and its applications- safety aspects.	12	CO5
	Total	60	
Course Outcomes			
Course Outcomes	On completion of this course, students will;		
CO1	Make use of the laboratory instruments- laminar air flow, pH meter, centrifugation methods, biosafety cabinets following SOP.	PO4, PO6, PO7, PO8, P11	
CO2	Apply chromatography techniques in the separation of biomolecules.	PO4, PO6, PO7, PO8, P11	
CO3	Perform molecular techniques like mutagenesis and their detection.	PO4, PO6, PO7, PO8, P11	
CO4	Estimate molecules in biological samples by adopting UV spectroscopic techniques.	PO4, PO6, PO7, PO8, P11	
CO5	Cultivate organisms anaerobically.	PO4, PO6, PO7, PO8, P11	

Text Books			
1.	Sharma B. K. (2014). Instrumental Method of Chemical Analysis. Krishna Prakashan Media (P) Ltd.		
2.	Chatwal G. R and Anand S. K. (2014.) Instrumental Methods of Chemical Analysis. Himalaya Publishing House.		
3.	Mitchell G. H. (2017). Gel Electrophoresis: Types, Applications and Research. Nova Science Publishers Inc.		
4.	Holme D. Peck H. (1998). Analytical Biochemistry. (3 rd Edition). Prentice Hall.		
5.	Jayaraman J. (2011). Laboratory Manual in Biochemistry. (2 nd Edition). Wiley Eastn Ltd., New Delhi.		
References Books			
1.	Pavia D. L. (2012) Spectroscopy (4 th Edition). Cengage.		
2.	Skoog A. and West M. (2014). Principles of Instrumental Analysis. (14 th Edition). W.B.Saunders Co., Philadelphia.		
3.	Miller J. M. (2007). Chromatography: Concepts and Contrasts (2 nd Edition) Wiley-Blackwell.		
4.	Gurumani N. (2006). Research Methodology for Biological Sciences. (1 st Edition) MJP Publishers.		
5.	Ponmurugan P. and Gangathara P. B. (2012). Biotechniques. (1 st Edition). MJP Publishers.		
Web Resources			
1.	https://norcaloa.com/BMIA		
2.	http://www.biologydiscussion.com/biochemistry/centrifugation/centrifuge-introduction- types-uses-and-other-details-with-diagram/12489		
3.	https://www.watelectrical.com/biosensors-types-its-working-and-applications .		
4.	http://www.wikiscales.com/articles/electronic-analytical-balance/		
5.	https://study.com/academy/lesson/what-is-chromatography-definition-types-uses .		
Methods of Evaluation			
Internal Evaluation	Continuous Internal Assessment Tests		25 Marks
	Assignments		
	Seminars		
	Attendance and Class Participation		
External Evaluation	End Semester Examination		75 Marks
	Total		100 Marks
Methods of Assessment			
Recall (K1)	Simple definitions, MCQ, Recall steps, Concept definitions		
Understand / Comprehend (K2)	MCQ, True/False, Short essays, Concept explanations, Short summary or overview		
Application (K3)	Suggest idea/concept with examples, Suggest formulae, Solve problems, Observe, Explain		
Analyse (K4)	Problem-solving questions, Finish a procedure in many steps, Differentiate between various ideas, Map knowledge		

Evaluate (K5)	Longer essay/ Evaluation essay, Critique or justify with pros and cons
Create (K6)	Check knowledge in specific or offbeat situations, Discussion, Debating or Presentations

Mapping with Programme Outcomes

	PO 1	PO 2	PO 3	PO 4	PO 5	PO 6	PO 7	PO 8	PO 9	PO 10	PO 11	PO 12	PO 13	PO 14
CO1				S		M	M	S			S			
CO2				S		M	M	S			S			
CO3				S		S	S	S			S			
CO4				S		M	S	S			S			
CO5				S		M	S	S			L			

Subject Code	Subject Name	Category	L	T	P	S	Credits	Inst. Hours	Marks		
									CIA	External	Total
22MBP GE2B	Herbal Technology and Cosmetic Microbiology	Elective Course II (Choice 2)	Y	Y	-	-	3	4	25	75	100
Course Objectives											
CO1	Impart knowledge of Indian Medicinal Plants and their applications in microbiology.										
CO2	Promote the technical skills involved in preparation of different types of plant extracts.										
CO3	Explain methods to analyze the antimicrobial activity of medicinal plants.										
CO4	Acquire knowledge on cosmetic microbiology and role of microorganisms in cosmetics.										
CO5	Gain insight into pharmacopeial microbial assays and biosafety.										
UNIT	Details								No. of Hours	Course Objectives	
I	Herbs, Herbal medicine - Indian medicinal plants: Scope and Applications of Indian medicinal plants in treating bacterial, fungal and viral diseases. Basic principles involved in Ayurvedha, Sidha, Unani and Homeopathy.								12	CO1	
II	Collection and authentication of selected Indian medicinal plants: <i>Emblica officinalis</i> , <i>Withania somnifera</i> , <i>Phyllanthus amarus</i> , <i>Tinospora cordifolia</i> , <i>Andrographis paniculata</i> , <i>Piper longum</i> , <i>Ocimum sanctum</i> , <i>Azardirchata indica</i> , <i>Terminalia chebula</i> , <i>Allium sativum</i> . Preparation of extracts- Hot and cold methods. Preparation of stock solutions.								12	CO2	
III	Antimicrobial activity of selected Indian medicinal Plants: - In vitro determination of antibacterial and fungal activity of selected whole medicinal plants/ parts – well-diffusion methods. MIC - Macro and micro dilution techniques. Antiviral activity- cell lines- cytotoxicity, cytopathic and non-cytopathic effect.								12	CO3	
IV	History of Cosmetic Microbiology – Need for cosmetic microbiology, Scope of cosmetic microbiology, - Role of microbes in cosmetic preparation. Preservation of cosmetics. Antimicrobial properties of natural cosmetic products – Garlic, neem, turmeric, aloe vera and tulsi. Sanitary practices in cosmetic manufacturing - HACCP protocols in cosmetic microbiology.								12	CO4	
V	Cosmetic microbiology test methods - Antimicrobial preservative efficacy, microbial content testing and biological toxicological testing. Validation methods - bioburden and Pharmacopeial microbial assays. Preservatives of cosmetics - Global regulatory and toxicological aspect of cosmetic preservatives.								12	CO5	
								Total	60		

Course Outcomes		
Course Outcomes	On completion of this course, students will;	
CO1	Identify the applications of Indian medicinal plants in treating diseases.	PO1, PO5
CO2	Identify and authenticate herbal plants.	PO6, PO7
CO3	Evaluate the antimicrobial activity of medicinal plants.	PO4, PO6, PO9
CO4	Describe the role of microorganisms and their metabolites in the preparation of cosmetics.	PO1, PO5, PO7
CO5	Validate procedures and biosafety measures in the mass production of cosmetics.	PO6, PO7
Text Books		
1.	Ayurvedic Formulary of India. (2011). Part 1, 2 & 3. Pharmacopoeia Commission for Indian Medicine and Homeopathy. ISBN-10:8190648977.	
2.	Panda H. (2004). Handbook on herbal medicines. Asia Pacific Business Press Inc. ISBN:8178330911.	
3.	Mehra P. S. (2019). A Textbook of Pharmaceutical Microbiology. Dreamtech Press. ISBN 13:9789389307344.	
4.	Geis P. A. (2020). Cosmetic microbiology: A Practical Approach. (3 rd Edition). CRC Press. ISBN:9780429113697.	
5.	Brannan D. K. (1997). Cosmetic microbiology: A Practical Handbook. CRC Press. ISBN-10:0849337135.	
References Books		
1.	Indian Herbal Pharmacopoeia (2002). Vol. I & II Indian Drug Manufacturers Association, Mumbai.	
2.	British Herbal Pharmacopoeia. (1990). Vol. I. British Herbal Medicine Association. ISBN: 0903032090.	
3.	Verpoorte R. and Mukherjee, P. K. (2010). GMP for Botanicals: Regulatory and Quality issues on Phytomedicines. In GMP for botanicals: regulatory and quality issues on phytomedicines. (2 nd edition). Saujanya Books, Delhi. ISBN-10:81-900788-5-2/8190078852. ISBN-13:978-81-900788-5-6/9788190078856.	
4.	Turner R. (2013). Screening methods in Pharmacology. Elsevier. ISBN:9781483264233.	
5.	Cupp M. J. (2010). Toxicology and Clinical Pharmacology of Herbal Products (pp. 85-93). M. J. Cupp. Humana Press. Totowa, NJ, USA. ISBN-10:1617371904.	
Web Resources		
1.	https://www.academia.edu/50236711/Modern_Extraction_Methods_for_Preparation_of_Bioactive_Plant_Extracts	
2.	https://www.nhp.gov.in/introduction-and-importance-of-medicinal-plants-and-herbs_mtl	
3.	https://pubmed.ncbi.nlm.nih.gov/17004305/	
4.	https://www.fda.gov/cosmetics/potential-contaminants-cosmetics/microbiological-safety-and-cosmetics	
5.	https://pubmed.ncbi.nlm.nih.gov/15156038/	

Methods of Evaluation			
Internal Evaluation	Continuous Internal Assessment Tests		25 Marks
	Assignments		
	Seminars		
	Attendance and Class Participation		
External Evaluation	End Semester Examination		75 Marks
	Total		100 Marks
Methods of Assessment			
Recall (K1)	Simple definitions, MCQ, Recall steps, Concept definitions		
Understand / Comprehend (K2)	MCQ, True/False, Short essays, Concept explanations, Short summary or overview		
Application (K3)	Suggest idea/concept with examples, Suggest formulae, Solve problems, Observe, Explain		
Analyse (K4)	Problem-solving questions, Finish a procedure in many steps, Differentiate between various ideas, Map knowledge		
Evaluate (K5)	Longer essay/ Evaluation essay, Critique or justify with pros and cons		
Create (K6)	Check knowledge in specific or offbeat situations, Discussion, Debating or Presentations		

Mapping with Programme Outcomes

	PO 1	PO 2	PO 3	PO 4	PO 5	PO 6	PO 7	PO 8	PO 9	PO 10	PO 11	PO 12	PO 13	PO 14
CO1	M				S									
CO2						S	M							
CO3				S		S			M					
CO4	M				S		S							
CO5						M	S							

Subject Code	Subject Name	Category	L	T	P	S	Credits	Inst. Hours	Marks		
									CIA	External	Total
22MBP GE2C	Essentials of Laboratory Management and Biosafety	Elective Course II (Choice 3)	Y	Y	-	-	3	4	25	75	100
Course Objectives											
CO1	To utilize containment principles to ensure biosafety.										
CO2	To enrich the student role and responsibilities of laboratory hazards and their control.										
CO3	To know the importance of first aid technique for various common lab accidents.										
CO4	To acquire knowledge of biosafety level, risk assessment and maintain proper hygiene in the laboratory.										
CO5	To discuss the biosafety regulations and guidelines and implementation of safety programs.										
UNIT	Details								No. of Hours	Course Objectives	
I	Introduction to the laboratory and laboratory hazards - General laboratory facilities – Occupational safety- Lab accidents - Fires, chemical burns, slips and falls, Animal bites. Cuts from broken glass. Toxic fume inhalation. General laboratory rules, Good laboratory practice (GLP). Laboratory plan.								12	CO1	
II	Common hazards in laboratory: Chemical hazards- Safe handling of chemicals and gases, hazard labels and symbols. Material safety datasheet (MSDS), Chemical handling - Fume hood, Storage of chemicals. Chemical Waste Disposal Guideline. Physical hazards - Physical agent data sheets (PADS), Electric hazards- Electrical shock, Electrical explosions, Electrical burns. Safe work practices. Potential ignition sources in the lab. Stages of Fire. Fire Extinguishers. Fire Response.								12	CO2	
III	Prevention and First aid for laboratory accidents. Personal protective equipment (PPE), Proper attire (Eye/Face Protection, laboratory coats, gloves, respirators. Disposal/Removal of PPE. Emergency equipment safety - Showers/ Eye Washes. Laboratory security and emergency response. First aid for - Injuries caused by broken glass, Acid/Alkali splashes on the skin, swallowing acid/alkali, burns caused by heat, electric shock.								12	CO3	
IV	Biosafety - Historical background. Blood borne pathogens (BBP) and laboratory - acquired infections. Introduction to biological safety cabinets. Primary containment for biohazards. Biosafety levels of specific microorganisms. Recommended biosafety. Levels for infectious agents and infected animals. Risk groups with examples - Risk assessment. Safety levels. Case studies - Safe working, hand hygiene. Laboratory instruments, packing, sending, transport, import and export of								12	CO4	

	biological agents. Hygiene, disinfection, decontamination, sterilization.		
V	Biosafety regulations and guidelines. Centers for disease control and prevention and the National institutes of health. Occupational safety and health administration. Recombinant DNA advisory committee(RDAC), Institutional biosafety committee(IBSC), Review committee on genetic manipulation(RCGM), Genetic engineering approval committee (GEAC). Implementation of biosafety guidelines.	12	CO5
	Total	60	
Course Outcomes			
Course Outcomes	On completion of this course, students will;		
CO1	Employ skills on laboratory safety and avoid laboratory accidents.	PO1, PO2, PO3, PO7, PO11	
CO2	Prevent laboratory hazards by practicing safety strategies.	PO2, PO5, PO7, PO11	
CO3	Practice various first aid procedures during common laboratory accidents.	PO1, PO2, PO3, PO5, PO10, PO11	
CO4	Ensure biosafety strategies in laboratory.	PO2, PO3, PO4, PO7, PO10, PO11	
CO5	Recognize the importance of biosafety guidelines.	PO3, PO4, PO5, PO7, PO10, PO11	
Text Books			
1.	Sateesh M. K. (2013). Bioethics and Biosafety, IK International Pvt Ltd. ISBN : 8190675702.		
2.	Muthuraj M. and Usharani B. (2019). Biosafety in Microbiological Laboratories. (1st Edition). Notion Press. ISBN 10: 1645878856		
3.	Biosafety in Microbiological and Biomedical Laboratories - U.S. Health Department and Human Services. (2016). (5 th Edition). Lulu.com.		
4.	Kanai. L. Mukherjee. (Medical Laboratory Technology(4 th Edition). CBS Publishers.		
5.	Ramakrishnan (2012). Manual of Medical Laboratory Techniques. JP brothers.		
References Books			
1.	World Health Organization, Biosafety programme management. (2010). (4 th Edition). WHO Publications.		
2.	Rashid N. (2013). Manual of Laboratory Safety (Chemical, Radioactive, and Biosafety with Biocides) (1 st Edition).		
3	Dayuan X. (2015). Biosafety and Regulation for Genetically Modified Organisms, Alpha Science International Ltd, ISBN-10 : 1842657917		
4.	Ochei J. Kolhatkar(2000). A. (Medical Laboratory Science – Theory and Practice. ISBN; 13:978-0074632239.		
5.	Lynne S. Garcia. Clinical Laboratory Management (2 nd Edition). ASM Press		
Web Resources			
1.	https://www.cdc.gov/labs/pdf/CDC-BiosafetymicrobiologicalBiomedicalLaboratories-2009-P.pdf		
2.	https://ucanapplym.s3.ap-south-1.amazonaws.com/RGU/notifications/E_learning/Online_study/PG-SEM-IV-Biosafety%20regulation.pdf		
3.	https://consteril.com/biosafety-levels-difference/		

4.	https://www.cdc.gov/labs/pdf/CDC-BiosafetymicrobiologicalBiomedicalLaboratories-2009-P.pdf	
5.	https://www.who.int/publications/i/item/9789240011311	
Methods of Evaluation		
Internal Evaluation	Continuous Internal Assessment Tests	25 Marks
	Assignments	
	Seminars	
	Attendance and Class Participitation	
External Evaluation	End Semester Examination	75 Marks
	Total	100 Marks
Methods of Assessment		
Recall (KI)	Simple definitions, MCQ, Recall steps, Concept definitions	
Understand / Comprehend (K2)	MCQ, True/False, Short essays, Concept explanations, Short summary or overview	
Application (K3)	Suggest idea/concept with examples, Suggest formulae, Solve problems, Observe, Explain	
Analyse (K4)	Problem-solving questions, Finish a procedure in many steps, Differentiate between various ideas, Map knowledge	
Evaluate (K5)	Longer essay/ Evaluation essay, Critique or justify with pros and cons	
Create (K6)	Check knowledge in specific or offbeat situations, Discussion, Debating or Presentations	

Mapping with Programme Outcomes

	PO 1	PO 2	PO 3	PO 4	PO 5	PO 6	PO 7	PO 8	PO 9	PO 10	PO 11	PO 12	PO 13	PO 14
CO1	S	S	S				S				S			
CO2		S			S		S				S			
CO3	S	S	S		S					S	S			
CO4		S	S	M			S			S	S			
CO5			S	S	S		S			S	S			

Subject Code	Subject Name	Category	L	T	P	S	Credits	Inst. Hours	Marks		
									CIA	External	Total
22MBPG CT4	Medical Bacteriology and Mycology	Core Course IV	Y	Y	-	-	4	6	25	75	100
Course Objectives											
CO1	Acquire Knowledge on collection, transportation and processing of various kinds of clinical specimens.										
CO2	Explain morphology, characteristics and pathogenesis of bacteria.										
CO3	Discuss various factors leading to pathogenesis of bacteria.										
CO4	Acquire knowledge on antifungal agents and their importance.										
CO5	Describe various diagnostic methods available for fungal disease diagnosis.										
UNIT	Details								No. of Hours	Course Objectives	
I	Classification of medically important bacteria, Normal flora of human body, Collection, transport, storage and processing of clinical specimens, Microbiological examination of clinical specimens, antimicrobial susceptibility testing.								20	CO1	
II	Morphology, classification, characteristics, pathogenesis, laboratory diagnosis and treatment of diseases caused by species of <i>Staphylococci</i> , <i>Streptococci</i> , <i>Pneumococci</i> , <i>Neisseriae.</i> , <i>Bacillus</i> , <i>Corynebacteria</i> , <i>Mycobacteria</i> and <i>Clostridium</i> .								20	CO2	
III	Morphology, classification, characteristics, pathogenesis, laboratory diagnosis and treatment of diseases caused by Enterobacteriaceae members, <i>Yersinia</i> , <i>Pseudomonas</i> , <i>Vibrio</i> , <i>Mycoplasma</i> , <i>Helicobacter</i> , <i>Rickettsiae</i> , <i>Chlamydiae</i> , <i>Bordetella</i> , <i>Francisella.</i> , <i>Spirochaetes-Leptospira</i> , <i>Treponema</i> and <i>Borrelia</i> . Nosocomial, zoonotic and opportunistic infections -prevention and control.								20	CO3	
IV	Morphology, taxonomy and classification of fungi. Detection and recovery of fungi from clinical specimens. Dermatophytes and agents of superficial mycoses. <i>Trichophyton</i> , <i>Epidermophyton</i> & <i>Microsporum</i> . Yeasts of medical importance – <i>Candida</i> , <i>Cryptococcus</i> . Mycotoxins. Antifungal agents.								15	CO4	
V	Dimorphic fungi causing Systemic mycoses, <i>Histoplasma</i> , <i>Coccidioides</i> , <i>Sporothrix</i> , <i>Blastomyces</i> . Fungi causing Eumycotic Mycetoma, Opportunistic fungi- Fungi causing secondary infections in immunocompromised patients. Immunodiagnostic methods in mycology.								15	CO5	
								Total	90		

Course Outcomes		
Course Outcomes	On completion of this course, students will;	
CO1	Collect, transport and process of various kinds of clinical specimens.	PO1,PO5,PO9
CO2	Analyze various bacteria based on morphology and pathogenesis.	PO1,PO5,PO9
CO3	Discuss various treatment methods for bacterial disease.	PO1,PO5,PO9
CO4	Employ various methods detect fungi in clinical samples and apply knowledge on antifungal agents..	PO5,PO9
CO5	Apply various immunodiagnostic method to detect fungal infections.	PO5,PO9
Text Books		
1.	Kanunga R. (2017). Ananthanarayanan and Panicker's Text book of Microbiology. (2017).Orient Longman, Hyderabad.	
2.	Greenwood, D., Slack, R. B. and Peutherer, J. F. (2012) Medical Microbiology, (18 th Edition). Churchill Livingstone, London.	
3.	Finegold, S. M. (2000) Diagnostic Microbiology, (10 th Edition). C.V. Mosby Company, St. Louis.	
4.	Alexopoulos C. J., Mims C. W. and Blackwell M. (2007). Introductory Mycology, (4 th Edition). Wiley Publishers.	
5.	Chander J. (2018). Textbook of Medical Mycology. (4 th Edition). Jaypee brothers Medical Publishers.	
References Books		
1.	Salle A. J. (2007). Fundamental Principles of Bacteriology. (4 th Edition). Tata McGraw-Hill Publications.	
2.	Collee J.C. Duguid J.P. Foraser, A.C, Marimon B.P, (1996). <u>Mackie & McCartney Practical Medical Microbiology</u> . 14 th edn, Churchill Livingston.	
3.	Cheesbrough M. (2006). <u>District Laboratory Practice in Tropical countries.- Part 22ndedn</u> .Cambridge University Press.	
4.	Topley and Wilson's. (1998). <u>Principles of Bacteriology</u> .9 th edn. Edward Arnold, London.	
5.	Murray P.R., Rosenthal K.S. and Michael A. (2013). <u>Medical Microbiology</u> . Pfaller. 7 th edn. Elsevier, Mosby Saunders.	
Web Resources		
1.	http://textbookofbacteriology.net/nd	
2.	https://microbiologysociety.org/members-outreach-resources/links.html	
3.	https://www.pathselective.com/micro-resources	
4.	http://mycology.cornell.edu/fteach.html	
5.	https://www.adelaide.edu.au/mycology/	

Methods of Evaluation		
Internal Evaluation	Continuous Internal Assessment Tests	25 Marks
	Assignments	
	Seminars	
	Attendance and Class Participation	
External Evaluation	End Semester Examination	75 Marks
	Total	100 Marks
Methods of Assessment		
Recall (K1)	Simple definitions, MCQ, Recall steps, Concept definitions	
Understand / Comprehend (K2)	MCQ, True/False, Short essays, Concept explanations, Short summary or overview	
Application (K3)	Suggest idea/concept with examples, Suggest formulae, Solve problems, Observe, Explain	
Analyze (K4)	Problem-solving questions, Finish a procedure in many steps, Differentiate between various ideas, Map knowledge	
Evaluate (K5)	Longer essay/ Evaluation essay, Critique or justify with pros and cons	
Create (K6)	Check knowledge in specific or offbeat situations, Discussion, Debating or Presentations	

Mapping with Programme Outcomes

	PO 1	PO 2	PO 3	PO 4	PO 5	PO 6	PO 7	PO 8	PO 9	PO 10	PO 11	PO 12	PO 13	PO 14
CO1	M				S				M					
CO2	M				S				M					
CO3	M				S				M					
CO4					S				M					
CO5					S				M					

Subject Code	Subject Name	Category	L	T	P	S	Credits	Inst. Hours	Marks		
									CIA	External	Total
22MBPGC T5	Medical Virology and Parasitology	Core Course V Theory	Y	Y	-	-	4	6	25	75	100
Course Objectives											
CO1	Describe the replication strategy and cultivation methods of viruses.										
CO2	Acquire knowledge about oncogenic virus and human viral infections.										
CO3	Develop diagnostic skills, in the identification of virus infections.										
CO4	Impart knowledge about parasitic infections.										
CO5	Develop diagnostic skills, in the identification of parasitic infections.										
UNIT	Details								No. of Hours	Course Objectives	
I	General properties of viruses - Structure and Classification - viroids, prions, satellite RNAs and virusoids. Cultivation of viruses - embryonated eggs, experimental animals and cell cultures. Purification and Assay of viruses – Physical and Chemical methods (Electron Microscopy, Protein and Nucleic acids studies.) Infectivity Assays (Plaque and end-point).								20	CO1	
II	Virus Entry, Host Defenses Against Viral Infections, Epidemiology, pathogenic mechanisms, Pathogenesis, laboratory diagnosis, treatment for the following viruses: DNA Viruses- Pox , Herpes , Adeno , Papova and Hepadna , RNA Viruses- Picorna, Orthomyxo, Paramyxo, Rhabdo, Rota, HIV and other Hepatitis viruses, Arbo – Dengue virus, Ebola virus, Emerging and reemerging viral infections								20	CO2	
III	Bacterial viruses - ΦX 174, M13, MU, T4, lambda, Pi; Structural organization, life cycle and phage production. Lysogenic cycle. Diagnosis of viral infections. Antiviral agents and viral vaccines.								15	CO3	
IV	Introduction to Medical Parasitology – Classification, host-parasite relationships. Epidemiology, life cycle, pathogenic mechanisms, laboratory diagnosis, treatment for the following: Protozoa causing human infections – <i>Entamoeba</i> , <i>Giardia</i> , <i>Trichomonas</i> , <i>Balantidium</i> , <i>Toxoplasma</i> , <i>Cryptosporidium</i> , <i>Leishmania</i> , and <i>Trypanasoma</i> .								15	CO4	
V	Classification, life cycle, pathogenicity, laboratory diagnosis and treatment for parasites – Helminthes - Cestodes – <i>Taenia Solium</i> , <i>T. Saginata</i> , <i>T.</i>								20	CO5	

	<i>Echinococcus</i> . Trematodes – <i>Fasciola Hepatica</i> , <i>Fasciolopsis Buski</i> , <i>Paragonimus</i> , <i>Schistosomes</i> . Nematodes - <i>Ascaris</i> , <i>Ankylostoma</i> , <i>Trichuris</i> , <i>Trichinella</i> , <i>Enterobius</i> , <i>Strongyloides</i> and <i>Wuchereria</i> . Other parasites causing infections in immune compromised hosts and AIDS.		
	Total	90	
Course Outcomes			
Course Outcomes	On completion of this course, students will;		
CO1	Cultivate viruses by different methods and aid in diagnosis. Perform purification and viral assay.	PO5, PO7, PO8, PO10	
CO2	Investigate the symptoms of viral infections and presumptively identify the viral disease.	PO5, PO7, PO8, PO10	
CO3	Diagnose various viral diseases by different methods.(serological, conventional and molecular)	PO5, PO7, PO8, PO10	
CO4	Educate public about the spread, control and prevention of parasitic diseases.	PO5, PO7, PO8, PO10	
CO5	Identify the protozoans and helminthes present in stool and blood specimens. Perform serological and molecular diagnosis of parasitic infections.	PO5, PO7, PO8, PO10	
Text Books			
1.	Kanunga R. (2017). Ananthanarayanan and Panicker’s Text book of Microbiology. (10 th Edition). Universities Press (India) Pvt. Ltd.		
2.	Dubey, R.C. and Maheshwari D.K. (2010). A Text Book of Microbiology. S. Chand & Co.		
3.	Rajan S. (2007). Medical Microbiology. MJP publisher.		
4.	Paniker J. (2006). Text Book of Parasitology. Jay Pee Brothers, New Delhi.		
5.	Arora, D. R. and Arora B. B. (2020). Medical Parasitology. (5 th Edition). CBS Publishers & Distributors Pvt. Ltd. New Delhi.		
Reference Books			
1.	Carter J. (2001). Virology: Principles and Applications (1 st Edition). Wiley Publications.		
2..	Willey J., Sandman K. and Wood D. Prescott’s Microbiology. (11 th Edition). McGraw Hill Book.		
3.	Jawetz E., Melnick J. L. and Adelberg E. A. (2000). Review of Medical Microbiology. (19 th Edition). Lange Medical Publications, U.S.A.		
4.	Finegold S.M. (2000). Diagnostic Microbiology. (10 th Edition). C.V. Mosby Company, St. Louis.		
5.	Levanthal R. and Cheadle R. S. (2012). Medical Parasitology. (6 th Edition). S.A. Davies Co. Philadelphia.		

Web Resources		
1.	https://en.wikipedia.org/wiki/Virology	
2.	https://academic.oup.com/femsre/article/30/3/321/546048	
3.	https://www.sciencedirect.com/science/article/pii/S0042682215000859	
4.	https://nptel.ac.in/courses/102/103/102103039/	
5.	https://www.healthline.com/health/viral-diseases#contagiousness	
Methods of Evaluation		
Internal Evaluation	Continuous Internal Assessment Tests	25 Marks
	Assignments	
	Seminars	
	Attendance and Class Participation	
External Evaluation	End Semester Examination	75 Marks
	Total	100 Marks

Methods of Assessment	
Recall (K1)	Simple definitions, MCQ, Recall steps, Concept definitions
Understand / Comprehend (K2)	MCQ, True/False, Short essays, Concept explanations, Short summary or overview
Application (K3)	Suggest idea/concept with examples, Suggest formulae, Solve problems, Observe, Explain
Analyses (K4)	Problem-solving questions, Finish a procedure in many steps, Differentiate between various ideas, Map knowledge
Evaluate (K5)	Longer essay/ Evaluation essay, Critique or justify with pros and cons
Create (K6)	Check knowledge in specific or offbeat situations, Discussion, Debating or Presentations

Mapping with Programme Outcomes

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO 10	PO 11	PO 12	PO 13	PO 14
CO1					M		L	L		M				
CO2					M		L	L		M				
CO3					M		L	L		M				
CO4					M		L	L		M				
CO5					M		L	L		M				

Subject Code	Subject Name	Category	L	T	P	S	Credits	Inst. Hours	Marks		
									CIA	External	Total
22MBPGCP 2	Practical II	Core Course VI-Practical I II	-	-	Y	-	4	6	25	75	100
Course Objectives											
CO1	Develop skills in the diagnosis of bacterial infections and antimicrobial sensitivity.										
CO2	Impart knowledge on fungal infections and its diagnosis.										
CO3	Diagnose parasitic										
CO4	To gain knowledge about industrially important microbes.										
CO5	Screen and utilize microorganisms for effective industrial production of metabolites.										
UNIT	Details							No. of Hours	Course Objectives		
I	Staining of clinical specimens - Wet mount, Differential and Special staining methods. Isolation and identification of bacterial pathogens from clinical specimens - cultivation in basal, differential, enriched, selective and special media – Biochemical identification tests.. Antimicrobial sensitivity testing - Kirby Bauer method and Stokes method. Minimum inhibitory concentration (MIC) test. Minimum bactericidal concentration (MBC) test.							20	CO1		
II	Identification and Classification of common fungi. Examination of different fungi by Lactophenol cotton blue staining. Examination of different fungi by KOH staining. Cultivation of fungi and their identification - <i>Mucor</i> , <i>Rhizopus</i> , <i>Aspergillus</i> , <i>Penicillium</i> . Isolation and characterization of bacteriophage from natural sources by phage titration. Cultivation of viruses –Egg Inoculation methods. Diagnosis of Viral Infections –ELISA –HIA. Spotters of viral inclusions and CPE-stained smears.							20	CO2		
III	Examination of parasites in clinical specimens - Ova/cysts in faeces. Concentration: methods – Flootation methods-simple Saturated salt solution method – Zinc							20	CO3		

	<p>sulphate methods - Sedimentation methods- Formal ether method. Blood smear examination for malarial parasites. Leishman's stain. Identification of common arthropods of medical importance - spotters of <i>Anopheles</i>, <i>Aedes</i>, Ticks and mites.</p>		
IV	<p>Good Laboratory Practices in Industrial Microbiology laboratory. Study of Bioreactor and its essential parts. Culturing and Characterization of microorganisms used in Dairy and Pharmaceutical industry. Screening for Enzyme producers (amylase /protease). Optimization of parameters for Amylase production. Screening for Organic acid producers (acetic acid/lactic acid). Screening for Antibiotic producers.</p>	15	CO4
V	<p>Immobilization of microbial cells and enzyme and its assessment. Microbiological assays of fermentation products – MIC- MBC. Microbiological assay of antibiotics by cup plate method and other methods.</p>	15	CO5
	Total	90	

Course Outcomes

Course Outcomes		On completion of this course, students will;
CO1	Collection of different clinical samples, transport, culture and examination.	PO7, PO8, PO9
CO2	Identify medically important bacteria, fungus and parasites from the clinical samples by staining and biochemical tests.	PO7, PO8, PO9
CO3	Promote diagnostic skills; interpret laboratory tests in the diagnosis of infectious diseases.	PO7, PO8, PO9, PO10
CO4	Perform antibiotic sensitivity tests and compare with the standard tests.	PO7, PO8, PO9, PO10

CO5	Screening of industrially important microbes for metabolite production.	PO7, PO8, PO9
Text Books		
1.	Cullimore D. R. (2010). Practical Atlas for Bacterial Identification, 2 nd Edition. Publisher-Taylor and Francis.	
2.	Abbott A.C. (2010). The Principles of Bacteriology. Nabu Press.	
3.	Parija S. C. (2012). Textbook of Practical Microbiology. Ahuja Publishing House.	
4.	Cappuccino, J. and Sherman, N. (2002) Microbiology: A Laboratory Manual, (6 th Edition). Pearson Education, Publication, New Delhi.	
5.	Morag C. and Timbury M.C. (1994). Medical Virology. 4 th edn. Blackwell Scientific Publishers.	
References Books		
1.	Collee J. G., Fraser A.G. Marmion B. P. and Simmons A. (1996). Mackie & McCartney Practical Medical Microbiology. (14 th Edition). Elsevier, New Delhi.	
2.	Chart H. (2018). Practical Laboratory Bacteriology. CRC Press.	
3.	Moore V. A. (2017). Laboratory Directions for Beginners in Bacteriology. Triste Publishing Ltd.	
4.	.Cheesbrough M. (2006). District Laboratory Practice in Tropical countries.- Part 22 nd Edition. Cambridge University Press.	
5.	Murray P.R., Rosenthal K.S. and Michael A. (2013). Medical Microbiology. Pfaller. 7 th Edition. Elsevier, Mosby Saunders	
Web Resources		
1.	http://textbookofbacteriology.net/	
2.	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7173454/	
3.	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3768729/	
4.	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC149666/	
5.	https://www.intechopen.com/books/current-issues-in-molecular-virology-viral-genetics-and-biotechnological-applications/vaccines-and-antiviral-agents	
Methods of Evaluation		
Internal Evaluation	Continuous Internal Assessment Tests	25 Marks
	Assignments	
	Seminars	
	Attendance and Class Participitation	
External Evaluation	End Semester Examination	75 Marks
	Total	100 Marks

Methods of Assessment	
Recall (K1)	Simple definitions, MCQ, Recall steps, Concept definitions
Understand / Comprehend (K2)	MCQ, True/False, Short essays, Concept explanations, Short summary or overview
Application (K3)	Suggest idea/concept with examples, Suggest formulae, Solve problems, Observe, Explain
Analyse (K4)	Problem-solving questions, Finish a procedure in many steps, Differentiate between various ideas, Map knowledge
Evaluate (K5)	Longer essay/ Evaluation essay, Critique or justify with pros and cons
Create (K6)	Check knowledge in specific or offbeat situations, Discussion, Debating or Presentations

Mapping with Programme Outcomes

	PO 1	PO 2	PO 3	PO 4	PO 5	PO 6	PO 7	PO 8	PO 9	PO 10	PO 11	PO 12	PO 13	PO 14
CO1							M	M	M					
CO2							M	M	M					
CO3							M	M	L	L				
CO4							M	M	M	L				
CO5							M	M	M					

Subject Code	Subject Name	Category	L	T	P	S	Credits	Inst. Hours	Marks		
									CIA	External	Total
22MBP GE3A	Epidemiology	Elective Course III (Choice 1)	Y	Y	-	-	3	4	25	75	100
Course Objectives											
CO1	Describe the role of epidemiology in public health.										
CO2	Explain about epidemiology tools and disease surveillance methods.										
CO3	Analyze various communicable and non-communicable diseases in India.										
CO4	Discuss on mechanism of antimicrobial resistance.										
CO5	Outline on National health programmes that have been designed to address the issues.										
UNIT	Details								No. of Hours	Course Objectives	
I	Fundamentals of epidemiology - Definitions of epidemiology – Epidemiology of infectious diseases in Public Health. Natural history of disease - Historical aspects of epidemiology. Common risk factors - Epidemiologic Triad - Agent factors, host factors and environmental factors. Transmission basics - Chain of infection, portal of entry. Modes of transmission -Direct and indirect. Stages of infectious diseases. Agents and vectors of communicable diseases of public health importance and dynamics of disease transmission. Epidemiology of Zoonosis - Control of zoonosis.								12	CO1	
II	Tools of Epidemiology - Measures of Disease - Prevalence, incidence. Index case. Risk rates. Descriptive Epidemiology - Cohort studies, measuring infectivity, survey methodology including census procedures. Surveillance strategies - Disease surveillance, geographical indication system, outbreak investigation in public health and contact investigation.								12	CO2	
III	Epidemiological aspects of diseases of national importance - Background to communicable and non-communicable diseases. Emerging disease threats - Severe Acute Respiratory Syndrome (SARS), Covid-19, Ebola, MDR-TB, Malaria, Mucor mycosis, Avian flu. Dengue, Swine Flu, Chikungunya. Epidemiology, prevention, and control of non-communicable diseases - Asthma, Coronary heart disease, Malignancy, diabetes mellitus, respiratory diseases, eye diseases, Dental disorders. Emerging and Re-emerging Diseases.								12	CO3	
IV	Mechanisms of Antimicrobial resistance - Multidrug Efflux pumps, Extended Spectrum β -lactamases (ESBL). Hospital acquired infections - Factors, infection sites, mechanisms, Role of Multidrug resistant pathogens. Role of <i>Pseudomonas</i> , <i>Acinetobacter</i> , <i>Clostridium difficile</i> , HBV, HCV, Rotavirus, <i>Cryptosporidium</i> and <i>Aspergillus</i> in Nosocomial infections. Prevention and management of nosocomial infections.								12	CO4	
V	National Programmes related to Communicable and Non-Communicable diseases - National Malaria Eradication Programme, Revised National Tuberculosis Control								12	CO5	

	Programme, Vector Borne Disease Control Programme, National AIDS Control Programme, National Cancer Control Programme and National Diabetes Control Programme.		
	Total	60	
Course Outcomes			
Course Outcomes	On completion of this course, students will;		
CO1	Apply the knowledge acquired on concepts of epidemiology to clinical and public health environment.		PO1
CO2	Plan various strategies to trace the epidemiology.		PO4, PO5, PO6
CO3	Plan the control of communicable and non-communicable diseases.		PO1, PO5,
CO4	Analyze the implications of drug resistance in the society and design the control of antimicrobial resistance and its management.		PO5,
CO5	Employ National control programs related to Communicable and Non-Communicable diseases with the public.		PO4, PO5,
Text Books			
1.	Dicker R., Coronado F., Koo. D. and Parrish. R. G. (2012). Principles of Epidemiology in Public Health Practice., (3 rd Edition). CDC.		
2.	Gerstman B. (2013). Epidemiology Kept Simple: An Introduction to Classic and Modern Epidemiology. (3 rd Edition). Wiley Blackwell.		
3.	Greenwood, D., Slack, R. B. and Peutherer, J. F. (2012) Medical Microbiology, (18 th Edition). Churchill Livingstone, London.		
4.	Jawetz E., Melnick J. L. and Adelberg E. A. (2000). Review of Medical Microbiology. (19 th Edition). Lange Medical Publications, U.S.A.		
5.	Dimmok N. J. and Primrose S. B. (1994). <u>Introduction to Modern Virology</u> .5 th edn. Blackwell Scientific Publishers.		
References Books			
1.	Bhopal R. S. (2016). Concepts of Epidemiology - An Integrated Introduction to the Ideas, Theories, Principles and Methods of Epidemiology. (3 rd Edition). Oxford University Press, New York.		
2.	Celentano D. D. and Szklo M. (2018). Gordis Epidemiology. (6 th Edition). Elseiver, USA.		
3.	Cheesbrough, M. (2004). District Laboratory Practice in Tropical Countries - Part 2, (2 nd Edition). Cambridge University Press.		
4.	Ryan K. J. and Ray C. G. (2004). Sherris Medical Microbiology. (4 th Edition), McGraw Hill, New York.		
5.	Topley W.W. C., Wilson, G. S., Parker M. T. and Collier L. H. (1998). Principles of Bacteriology. (9 th Edition). Edward Arnold, London.		
Web Resources			
1.	https://www.scielo.br/j/rbca/a/mjDFGTtfWtBm786ZmR9TG9d/?lang=en		
2.	https://hal.archives-ouvertes.fr/hal-00902711/document		
3.	https://www.who.int/csr/resources/publications/whocdscsreph200212.pdf		
4.	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7187955/		

Subject Code	Subject Name	Category	L	T	P	S	Credits	Inst. Hours	Marks		
									CIA	External	Total
22MBPG E3B	Clinical and Diagnostic Microbiology	Elective Course III (Choice 2)	Y	Y	-	-	3	4	25	75	100
Course Objectives											
CO1	Describe appropriate safety protocol and laboratory techniques for handling specimens and biomedical waste management.										
CO2	Develop working knowledge of techniques used to identify infectious agents in the clinical microbiology lab.										
CO3	Elucidate various diagnostic procedures in microbiology.										
CO4	Acquire knowledge on different methods employed to check antibiotic sensitivity.										
CO5	Gain knowledge on hospital acquired infections and their control measures.										
UNIT	Details								No. of Hours	Course Objectives	
I	Microbiology Laboratory Safety Practices -General Safety Guidelines, Handling of Biological Hazards, Infectious health care waste disposal - Biomedical waste management, Emerging and Re-emerging infections.								12	CO1	
II	Diagnostic procedures - General concept of Clinical specimen collection, transport, storage and general processing in Microbiology laboratory - Specimen acceptance and rejection criteria.								12	CO2	
III	Diagnosis of microbial diseases - Microbiological, immunological and molecular diagnosis of microbial diseases. Automation in Microbial diagnosis.								12	CO3	
IV	Antibiotic sensitivity tests - Disc diffusion - Stokes and Kirby Bauer methods- Dilution - MBC/MIC - Quality control for antibiotics and standard strains.								12	CO4	
V	Nosocomial infections – common types, sources, reservoir and mode of transmission, pathogenesis and control measures. Hospital Infection Control Committee (HICC) – Functions.								12	CO5	
	Total								60		

Course Outcomes		
Course Outcomes	On completion of this course, students will;	
CO1	Apply Laboratory safety procedures and hospital waste disposal strategies.	PO5, PO6, PO7
CO2	Collect various clinical specimens, handle, preserve and process safely.	PO6, PO7
CO3	Identify the causative agents of diseases by conventional and molecular methods following standard protocols.	PO6, PO7, PO9, PO11
CO4	Assess the antimicrobial susceptibility pattern of pathogens.	PO7, PO9
CO5	Trace the sources of nosocomial infection and recommend control measures.	PO5, PO7
TEXT BOOKS		
1.	Collee J. G., Fraser A.G. Marmion B. P. and Simmons A. (1996). Mackie & McCartney Practical Medical Microbiology. (14 th Edition). Elsevier, New Delhi. ISBN-10:0443047219 / ISBN-13-978-0443047213.	
2.	Tille P. M. (2021). Bailey and Scott's Diagnostic Microbiology. (15 th Edition). Elsevier. ISBN:9780323681056.	
3.	Jawetz E., Melnick J. L. and Adelberg E. A. (2000). Review of Medical Microbiology. (19 th Edition). Lange Medical Publications, U.S.A.	
4.	Mukherjee K.L. (2000). Medical Laboratory Technology. Vol. 1-3. (2 nd Edition). Tata McGraw-Hill Education. ISBN-10:0074632604.	
5.	Sood R. (2009). Medical Laboratory Technology – Methods and Interpretations. (6 th Edition). Jaypee Brothers Medical Publishers (P) Ltd. New Delhi. ISBN:9788184484496.	
References Books		
1.	Murray P. R., Baron E. J., Jorgenson J. H., Pfaller M. A. and Tenover F.C. (2003). Manual of Clinical Microbiology. (8 th Edition). American Society for Microbiology, Washington, DC. ISBN:1-555810255-4.	
2.	Bennett J. E., Dolin R. and Blaser M. J. (2019). Principles and Practice of Infectious Diseases. (9 th Edition). Elsevier. EBook ISBN:9780323550277. Hardcover ISBN:9780323482554.	
3.	Ridgway G. L., Stokes E. J. and Wren M. W. D. (1987). Clinical Microbiology 7 th Edition. Hodder Arnold Publication. ISBN-10:0340554231 / ISBN-13:9780340554234.	
4.	Koneman E.W., Allen S. D., Schreckenber P. C. and Winn W. C. (2020). Koneman's Color Atlas and Textbook of Diagnostic Microbiology. (7 th Edition). Jones & Bartlett Learning. ISBN:1284322378 9781284322378.	
5.	Cheesbrough, M. (2004). District Laboratory Practice in Tropical Countries - Part 2, (2 nd Edition). Cambridge University Press. ISBN-13:978-0-521-67631-1 / ISBN-10:0-521-67631-2.	
Web Resources		
1.	https://www.ncbi.nlm.nih.gov/books/NBK20370/	
2.	https://www.msdmanuals.com/en-in/home/infections/diagnosis-of-infectious3disease/diagnosis-of-infectious-disease	

3.	https://journals.asm.org/doi/10.1128/JCM.02592-20
4.	https://www.sciencedirect.com/science/article/pii/S2221169116309509
5.	http://www.textbookofbacteriology.net/normalflora_3.html

Methods of Evaluation

Internal Evaluation	Continuous Internal Assessment Tests	25 Marks
	Assignments	
	Seminars	
	Attendance and Class Participation	
External Evaluation	End Semester Examination	75 Marks
	Total	100 Marks

Methods of Assessment

Recall (K1)	Simple definitions, MCQ, Recall steps, Concept definitions
Understand / Comprehend (K2)	MCQ, True/False, Short essays, Concept explanations, Short summary or overview
Application (K3)	Suggest idea/concept with examples, Suggest formulae, Solve problems, Observe, Explain
Analyze (K4)	Problem-solving questions, Finish a procedure in many steps, Differentiate between various ideas, Map knowledge
Evaluate (K5)	Longer essay/ Evaluation essay, Critique or justify with pros and cons
Create (K6)	Check knowledge in specific or offbeat situations, Discussion, Debating or Presentations

Mapping with Programme Outcomes

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO 10	PO 11	PO 12	PO 13	PO 14
CO1					S	M	M							
CO2						M	S							
CO3						M	S		M		S			
CO4							S		M					
CO5					S		M							

Subject Code	Subject Name	Category	L	T	P	S	Credits	Inst. Hours	Marks		
									CIA	External	Total
22MBPG E3C	Bioremediation	Elective Course III (Choice 3)	Y	Y	-	-	3	4	25	75	100
Course Objectives											
CO1	Describe the nature and importance of bioremediation and use in real world applications.										
CO2	Describe the typical composition of waste water and application of efficient technologies for water treatment.										
CO3	Explain the fundamentals of treatment technologies and the considerations for its design and implementation in treatment plants.										
CO4	Explain the potential of microbes in ore extraction and acquaint students with methods of reducing health risks caused by xenobiotics.										
CO5	Familiarize the role of plants and their associated microbes in remediation and management of environmental pollution.										
UNIT	Details								No. of Hours	Course Objectives	
I	Bioremediation - process and organisms involved. Bioaugmentation - Ex-situ and in-situ processes; Intrinsic and engineered bioremediation. Major pollutants and associated risks; organic pollutant degradation. Microbial aspects and metabolic aspects. Factors affecting the process. Recent developments and significance.								12	CO1	
II	Microbes involved in aerobic and anaerobic processes in nature. Water treatment, BOD, COD, dissolved gases, removal of heavy metals, total organic carbon removal. Secondary waste water treatments- use of membrane bioreactor.								12	CO2	
III	Composting of solid wastes, anaerobic digestion - methane production and important factors involved, Pros and cons of anaerobic process, sulphur, iron and nitrate reduction, hydrocarbon degradation, degradation of nitroaromatic compounds. Bioremediation of dyes, bioremediation in paper and pulp industries.								12	CO3	
IV	Microbial leaching of ores - process, microorganisms involved and metal recovery with special reference to copper and iron. Biotransformation of heavy metals and xenobiotics. Petroleum biodegradation - reductive and oxidative. Dechlorination. Biodegradable of plastics and super bug.								12	CO4	
V	Phytoremediation of heavy metals in soil - Basic principles of phytoremediation - Uptake and transport, Accumulation and sequestration. Phytoextraction. Phytodegradation. Phytovolatilization. Rhizodegradation. Phytostabilization – Organic and synthetic amendments in multi metal contaminated mine sites. Role of Arbuscular mycorrhizal fungi and plant growth promoting rhizobacteria in phytoremediation.								12	CO5	
Total								60			

Course Outcomes		
Course Outcomes		
CO1	Differentiate Ex-situ bioremediation and In-situ bioremediation. Assess the roles of organisms in bioremediation.	PO1, PO2, PO4, PO5
CO2	Distinguish microbial processes necessary for the design and optimization of biological processing unit operations.	PO1, PO4, PO5, PO11
CO3	Identify, formulate and design engineered solutions to environmental problems.	PO5, PO7, PO8, PO11
CO4	Explore microbes in degradation of toxic wastes and playing role on biological mechanisms.	PO5, PO6, PO7, PO8, PO9
CO5	Establish the mechanisms of Arbuscular mycorrhizal fungi and Plant growth promoting <i>Rhizobacteria</i> in phytoremediation.	PO1, PO5, PO6, PO7, PO8
Text Books		
1.	Bhatia H.S. (2018). A Text book on Environmental Pollution and Control. (2 nd Edition). Galgotia Publications.	
2.	Chatterjee A. K. (2011). Introduction to Environmental Biotechnology. (3 rd Edition). Printice-Hall, India.	
3.	Pichtel, J. (2014). Waste Management Practices: Municipal, Hazardous, and Industrial, 2 nd edition, CRC Press.	
4.	Liu, D.H.F and Liptak, B.G (2005). Hazardous Wastes and Solid Wastes, Lewis Publishers.	
5.	Rajendran, P. & Gunasekaran, P. (2006). Microbial Bioremediation. 1 st edition. MJP Publishers	
References Books		
1.	Sangeetha J., Thangadurai D., David M. and Abdullah M.A. (2016). Environmental Biotechnology: Biodegradation, Bioremediation, and Bioconversion of Xenobiotics for Sustainable Development. (1 st Edition). Apple Academic Press.	
2.	Singh A. and Ward O. P. (2004). Biodegradation and Bioremediation. Soil Biology. Springer.	
3.	Singh A., Kuhad R. C., and Ward O. P. (2009). Advances in Applied Bioremediation (1 st Edition). Springer-Verlag Berlin Heidelberg, Germany.	
4.	Atlas, R.M & Bartha, R. (2000). Microbial Ecology. Addison Wesley Longman Inc.	
5.	Rathoure, A.K. (Ed.). (2017). Bioremediation: Current Research and Applications. 1 st edition. I.K. International Publishing House Pvt. Ltd.	
Web Resources		
1.	Bioremediation- Objective, Principle, Categories, Types, Methods, Applications (microbenotes.com)	
2.	https://agris.fao.org > agris-search	
3.	https://www.sciencedirect.com/topics/earth-and-planetary-sciences/bioremediation	
4.	https://www.intechopen.com/chapters/70661	

5.	https://microbiologysociety.org/blog/bioremediation-the-pollution-solution.html	
Methods of Evaluation		
Internal Evaluation	Continuous Internal Assessment Tests	
	Assignments	
	Seminars	
	Attendance and Class Participation	
External Evaluation	End Semester Examination	75 Marks
Total		100 Marks
Methods of Assessment		
Recall (K1)	Simple definitions, MCQ, Recall steps, Concept definitions	
Understand / Comprehend (K2)	MCQ, True/False, Short essays, Concept explanations, Short summary or overview	
Application (K3)	Suggest idea/concept with examples, Suggest formulae, Solve problems, Observe, Explain	
Analyse (K4)	Problem-solving questions, Finish a procedure in many steps, Differentiate between various ideas, Map knowledge	
Evaluate (K5)	Longer essay/ Evaluation essay, Critique or justify with pros and cons	
Create (K6)	Check knowledge in specific or offbeat situations, Discussion, Debating or Presentations	

Mapping with Programme Outcomes

	PO 1	PO 2	PO 3	PO 4	PO 5	PO 6	PO 7	PO 8	PO 9	PO 10	PO 11	PO 12	PO 13	PO 14
CO1	S	M		M	S									
CO2	S			M	S						S			
CO3					S		S	S			S			
CO4					S	S	S	S	S					
CO5	M				S	M	S	S						

Subject Code	Subject Name	Category	L	T	P	S	Credits	Inst. Hours	Marks		
									CIA	External	Total
22MBP GE4A	Bioinformatics	Elective Course IV Theory (Choice 1)	Y	Y	-	-	3	4	25	75	100

Course Objectives

CO1	Discuss about various biological data mining concepts, tools.
CO2	Elucidate the principles and applications of sequence alignment methods and tools.
CO3	Demonstrate different phylogenetic tree construction methods and its uses in phylogenetic analysis.
CO4	Acquaint with various approaches in predicting 3D and 2D structure of proteins.
CO5	Describe various tools and techniques used in molecular docking, immunoinformatics and subtractive genomics.

UNIT	Details	No. of Hours	Course Objectives
I	Biological Data Mining – Exploration of Data Mining Tools. Cluster Analysis Methods. Data Visualization. Biological Data Management. Biological Algorithms – Biological Primary and Derived Databases.	12	CO1
II	Phylogenetic Tree Construction - Concept of Dendrograms. Evolutionary Trees - Distance Based Tree Reconstruction - Ultrametric trees and Ultrametric distances – Reconstructing Trees from Additive Matrices - Evolutionary Trees and Hierarchical Clustering - Character Based Tree Reconstruction - Maximum Parsimony Method, Maximum likelihood method - Reliability of Trees – Substitution matrices – Evolutionary models.	12	CO2
III	Computational Protein Structure prediction – Secondary structure – Homology modelling- Fold recognition and ab initio 3D structure prediction – Structure comparison and alignment – Prediction of function from structure.	12	CO3
IV	Prediction of Properties of Ligand Compounds – 3D Autocorrelation –Prediction of the Toxicity of Compounds	12	CO4
V	Molecular Docking- Flexible - Rigid docking- Target- Ligand preparation- Solvent accessibility- Surface volume calculation, Active site prediction- Docking algorithms- Genetic, Lamarckian - Docking analyses- Molecular interactions, bonded and nonbonded - Molecular Docking Software and Working Methods. Genome to drug discovery – Subtractive Genomics – Principles of Immunoinformatics and Vaccine Development.	12	CO5
	Total	60	

Course Outcomes		
Course Outcomes	On completion of this course, students will;	
CO1	Access to databases that provides information on nucleic acids and proteins.	PO1, PO4, PO6, PO7, PO9, PO10, PO13
CO2	Invent algorithms for sequence alignment.	PO7, PO9, PO10, PO13
CO3	Construct phylogenetic tree.	PO6, PO9, PO10
CO4	Predict the structure of proteins.	PO4, PO6, PO7, PO9, PO13
CO5	Design drugs by predicting drug ligand interactions and molecular docking.	PO4, PO5, PO6, PO7, PO9, PO10, PO13
Text Books		
1.	Lesk A. M. (2002). Introduction to Bioinformatics. (4 th Edition). Oxford University Press.	
2.	Lengauer T. (2008). Bioinformatics- from Genomes to Therapies (Vol-1).Wiley- VCH.	
3.	Rastogi S. C., Mendiratta N. and Rastogi P. (2014). Bioinformatics - Methods and Applications (Genomics, Proteomics and Drug Discovery) (4 th Edition). Prentice-Hall of India Pvt.Ltd.	
4.	Attwood, T.K. and Parry-Smith, D.J. (1999). Introduction to Bioinformatics. Addison Wesley Longman Limited, England.	
5.	Mount D.W., (2013).Bioinformatics sequence and genome analysis, 2 nd edn.CBS Publishers, New Delhi.	
References Books		
1.	Baxevanis A. D. and Ouellette F. (2004). Bioinformatics: A Practical Guide to the Analysis of Genes and Proteins. (2nd Edition). John Wiley and Sons.	
2.	Bosu O. and Kaur S. (2007). Bioinformatics - Database, Tools, and Algorithms. Oxford University Press.	
3.	David W. M. (2001). Bioinformatics Sequence and Genome Analysis (2 nd Edition). CBS Publishers and Distributors(Pvt.)Ltd.	
4.	Xiong J, (2011). <u>Essential bioinformatics</u> , First south Indian Edition, Cambridge University Press.	
5.	Harshawardhan P.Bal, (2006). <u>Bioinformatics Principles and Applications</u> , Tata McGraw-Hill Publishing Company Limited.	
Web Resources		
1.	https://www.hsls.pitt.edu/obrc/	
2.	https://www.hsls.pitt.edu/obrc/index.php?page=dna	
3.	https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1669712/	
4.	https://www.ebi.ac.uk/	
5.	https://www.kegg.jp/kegg/kegg2.html	

Methods of Evaluation			
Internal Evaluation	Continuous Internal Assessment Tests		25 Marks
	Assignments		
	Seminars		
	Attendance and Class Participation		
External Evaluation	End Semester Examination		75 Marks
	Total		100 Marks
Methods of Assessment			
Recall (KI)	Simple definitions, MCQ, Recall steps, Concept definitions		
Understand / Comprehend (K2)	MCQ, True/False, Short essays, Concept explanations, Short summary or overview		
Application (K3)	Suggest idea/concept with examples, Suggest formulae, Solve problems, Observe, Explain		
Analyse (K4)	Problem-solving questions, Finish a procedure in many steps, Differentiate between various ideas, Map knowledge		
Evaluate (K5)	Longer essay/ Evaluation essay, Critique or justify with pros and cons		
Create (K6)	Check knowledge in specific or offbeat situations, Discussion, Debating or Presentations		

Mapping with Programme Outcomes

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO 10	PO 11	PO 12	PO 13	PO 14
CO1	M			M		M			M	M			M	
CO2							S		S	S			S	
CO3						S			S	S				
CO4				S		S	S		S				S	
CO5				S	S	S	S		S	S			S	

Subject Code	Subject Name	Category	L	T	P	S	Credits	Inst. Hours	Marks		
									CIA	External	Total
22MBP GE4B	Nanobiotechnology	Elective Course IV (Choice 2)	Y	Y	-	-	3	4	25	75	100
Course Objectives											
CO1	Analyze nanomaterials based on the understanding of nanobiotechnology.										
CO2	Discuss the methods of fabrication of nanomaterials.										
CO3	Gain Knowledge on characterization of nanomaterials.										
CO4	Discover nanomaterials for targeted drug delivery.										
CO5	Explain nanomaterials in nanomedicine and environmental pollution.										
UNIT	Details								No. of Hours	Course Objectives	
I	Introduction to nanobiotechnology, Nano size-changing phenomena at nano scale, Classification of nanomaterials based on their dimensions (0D, 1D, 2D and 3D materials) and based on realization of their applications (The First, second, third and fourth generation materials), Class of nanomaterials and their applications. Need for nanomaterials and the risks associated with the materials.								12	CO1	
II	Fabrication of Nanomaterials-Top-down and Bottom-up approaches, Solid phase synthesis-milling, Liquid phase synthesis-Sol-gel synthesis, colloidal synthesis, micro emulsion method, hydrothermal synthesis and solvo thermal synthesis, Vapour/Gas phase synthesis-Inert gas condensation, flame pyrolysis, Laser ablation and plasma synthesis techniques. Microbial synthesis of nanoparticles.								12	CO2	
III	Characterization of nanoparticles – Based on particle size/morphology- Dynamic light scattering (DLS), Scanning electron microscopy (SEM), Transmission electron microscopy (TEM), Atomic force microscopy (AFM), Based on surface charge-zeta potential, Based on structure –X-ray diffraction (XRD), Fourier transform infrared spectroscopy (FTIR), Energy dispersive X-ray analysis (EDX), Based on optical properties- UV – Spectrophotometer, Based on magnetic properties-Vibrating sample magnetometer (VSM).								12	CO3	
IV	Nanomaterial based Drug delivery and therapeutics-surface modified nano particles, MEMS/NEMS based devices, peptide/DNA coupled nanoparticles, lipid and inorganic nano particles for drug delivery, Metal/metaloxide nano particles as antibacterial, antifungal and antiviral agents. Toxicity of nanoparticles and Toxicity Evaluation.								12	CO4	
V	Nanomaterials in diagnosis-Imaging, nanosensors in detection of pathogens. Treatment of surface water, ground								12	CO5	

	water and waste water contaminated by toxic metal ions, organic and inorganic solutes and microorganisms.		
	Total	60	
Course Outcomes			
Course Outcomes	On completion of this course, students will;		
CO1	Employ knowledge in the field of nanobiotechnology for development.	PO1, PO9	
CO2	Identify various applications of nanomaterials in the field of medicine and environment.	PO1, PO9	
CO3	Examine the prospects and significance of nanobiotechnology.	PO1, PO6, PO11	
CO4	Identify recent advances in this area and create a career or pursue research in the field.	PO1, PO5, PO7, PO9	
CO5	Design non-toxic nanoparticles for targeted drug delivery.	PO1, PO5, PO7, PO9, PO11	
Text Books			
1.	Brydson R. M., Hammond, C. (2005). Generic Methodologies for Nanotechnology: Characterization. In Nanoscale Science and Technology. John Wiley & Sons, Ltd.		
2.	Leggett G. J., Jones R. A. L. (2005). Bionanotechnology. In Nanoscale Science and Technology. John Wiley & Sons, Ltd.		
3.	Mohan Kumar G. (2016). Nanotechnology: Nanomaterials and nanodevices. Narosa Publishing House.		
4.	Goodsell D. S. (2004). Bionanotechnology. John Wiley & Sons, Inc.		
5.	Pradeep T. (2007). Nano: The Essentials-Understanding nanoscience and nanotechnology. Tata McGraw-Hill.		
References Books			
1.	Nouailhat A. (2008). An Introduction to Nanoscience and Nanotechnology, Wiley.		
2.	Sharon M. and Maheshwar (2012). Bio-Nanotechnology: Concepts and Applications. New Delhi. Ane books Pvt Ltd.		
3.	Niemeyer C.M. and Mirkin C. A. (2005). Nanobiotechnology. Wiley Interscience.		
4.	Rehm, B. (2006). Microbial Bionanotechnology: Biological Self-Assembly Systems and Biopolymer-Based Nanostructures. Horizon Scientific Press.		
5..	Reisner, D.E. (2009). Bionanotechnology: Global Prospects. CRC Press		
Web Resources			
1.	https://www.gale.com/nanotechnology		
2.	https://www.understandingnano.com/resources.html		
3.	http://dbtnanobiotech.com/index2.php		
4.	http://www.istl.org/11-winter/internet1.html		
5.	https://www.cdc.gov/niosh/topics/nanotech/default.html		

Methods of Evaluation			
Internal Evaluation	Continuous Internal Assessment Tests		25 Marks
	Assignments		
	Seminars		
	Attendance and Class Participation		
External Evaluation	End Semester Examination		75 Marks
	Total		100 Marks
Methods of Assessment			
Recall (K1)	Simple definitions, MCQ, Recall steps, Concept definitions		
Understand/Comprehended (K2)	MCQ, True/False, Short essays, Concept explanations, Short summary or overview		
Application (K3)	Suggest idea/concept with examples, Suggest formulae, Solve problems, Observe, Explain		
Analyse (K4)	Problem-solving questions, Finish a procedure in many steps, Differentiate between various ideas, Map knowledge		
Evaluate (K5)	Longer essay/ Evaluation essay, Critique or justify with pros and cons		
Create (K6)	Check knowledge in specific or offbeat situations, Discussion, Debating or Presentations		

Mapping with Programme Outcomes

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO10	PO11	PO12	PO13	PO14
CO1	S			M					M					
CO2	S								S					
CO3	S					M					S			
CO4	S				S		M		S					
CO5	S				S		M		S		S			

Subject Code	Subject Name	Category	L	T	P	S	Credits	Inst. Hours	Marks		
									CIA	External	Total
22MBPG E4C	Clinical Research And Clinical Trials	Elective Course IV (Choice 3)	Y	Y	-	-	3	4	25	75	100
Course Objectives											
CO1	Provide an overview of history and methods involved in conducting clinical research.										
CO2	Design the principles involved in ethical, legal, and regulatory issues in clinical research on human subjects.										
CO3	Describe principles and issues involved in monitoring patient-oriented research.										
CO4	Formulate a well- defined quality assurance and quality control plans.										
CO5	Acquire business development skills in the area of clinical research.										
UNIT	Details								No. of Hours	Course Objectives	
I	Introduction to Clinical Research: Clinical Research: An Overview, Different types of Clinical Research. Clinical Pharmacology: Pharmacokinetics, Pharmacodynamics, Pharmacoepidemiology, Bioavailability, Bioequivalence, Terminologies and definition in Clinical Research. Drug Development Process: Drug Discovery Pipeline, Drug Discovery Process. Preclinical trail, Human Pharmacology (Phase-I), Therapeutic Exploratory trail (Phase-II), Therapeutic Confirmatory Trail (Phase-III) and Post marketing surveillance (Phase-IV).								12	CO1	
II	Ethical Considerations and Guideline in Clinical Research: Historical guidelines in Clinical Research-Nuremberg code, Declaration of Helsinki, Belmont report. International Conference on Harmonization (ICH)-Brief history of ICH, Structure of ICH & ICH Harmonization Process, Guidelines for Good Clinical Practice. Regulation in Clinical Research- Drug and cosmetic act, FDA, Schedule-Y- Ethics Committee and their responsibilities. Clinical Research Regulatory Submission & approval Process- IND, NDA and ANDA submission Procedure. DCGI submission procedure. Other Regulatory authorities- EMEA, MHRA, PhRMA.								12	CO2	
III	Clinical Trial Management: Key Stakeholders in Clinical Research, Ethics Committees and Institutional Review Board, Responsibilities of Sponsor. Responsibilities of Investigator, Protocol in Clinical Research Clinical Trial Design, Project Planning Project Managements - Informed Consent, Investigator's Brochure (IB), Selection of an Investigator and Site, Patient screening, Inclusion and exclusion criteria, Randomization, Blinding. Essential Documents in clinical research -IB, ICF, PIS, TMF, ISF, CDA & CTA.								12	CO3	
IV	Quality Assurance, Quality Control & Clinical Monitoring: Defining the terminology-Quality, Quality system, Quality Assurance & Quality Control-QA audit plan. 21 CFR Part 11, Site Auditing, Sponsor Compliance and Auditing, SOP For								12	CO4	

	Clinical Research-CRF Review & Source Data Verification, Drug Safety Reporting Corrective and preventative action process.		
V	Business Development in the Clinical Research Industry: Introduction & Stages of Business Development-Start-up Phase, Growth Phase, Maturity Phase, Decline Phase. Outsourcing in Clinical Research, Reasons for outsourcing to contract research organizations, The India Advantage, Scope and Future of CRO, List of Clinical Research Organizations in India, List of IT companies offering services in Clinical Research. Role of business development manager.	12	CO5
	Total	60	
Course Outcomes			
Course Outcomes	On completion of this course, students will;		
CO1	Apprehend the Drug Development process and different phases of clinical trials.	PO1, PO2, PO3, PO5	
CO2	Recognize the ethics and regulatory perspectives on clinical research trials activities.	PO3, PO5, PO6, PO9	
CO3	Accentuate about clinical trials management concepts and documentation process.	PO2, PO4, PO6, PO9	
CO4	Accomplish quality assurance and quality control to ensure the protection of human subjects and the reliability of clinical trial results.	PO2, PO4, PO6, PO7, PO9	
CO5	To nurture skills recitation to commercial start up and industriousness.	PO4, PO8, PO9, PO11, PO13	
Text Books			
1.	Gallin J. I., Ognibene F. P. and Johnson L. L. (2007). Principles and Practice of Clinical Research. (4 th Edition). Elsevier, 2007.ISBN-10: 0128499052		
2.	Friedman L. M., Furberg C. D. and Demets D. (1998). Fundamentals of Clinical Trials, Vol: XVIII. (3 rd Edition). Springer Science & Business Media.		
3.	Hulley S. B., Cummings S. R., Browner W. S., Grady D. G. and Newman T. B. (2013). Designing Clinical Research. (4 th Edition). Jaypee Medical. ISBN-13: 978-1608318049.		
4.	Reed,G. (2004). Prescott and Dunn's Industrial Microbiology, 4 th edn, CBS publication and distributors.		
5.	Himanshu B. Text book of Clinical Research, Pee Vee books.		
References Books			
1.	Friedman L.M., Fuberge C.D., DeMets D. and Reboussen, D.M. (2015). Fundamentals of Clinical Trials, Springer.		
2.	Browner W. S., (2012). Publishing and Presenting Clinical Research. (3 rd Edition). Lippincott Williams and Wilkins.		
3.	Rondel R. K., Varley S. A. and Webb C. F. (2008). Clinical Data Management. (2 nd Edition). Wiley.		
4.	Peppler, H.J. and Pearl Man, D. (1979). Fermentation Technology, Vol 1 & 2, 2 nd Edition Academic Press, London.		

5.	E1-Mansi, E.M.T., Bryce, C.F.A., Demain, A.L. and Allman,A.R. (2007). Fermentation Microbiology and Biotechnology. 2 nd Edition, CRC press, Taylor and Francis Group.
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Web Resources

1	https://www.hzu.edu.in/uploads/2020/10/Textbook-of-Clinical-Trials-Wiley-(2004).pdf
2	https://www.routledge.com/A-Practical-Guide-to-Managing-Clinical-Trials/Pfeiffer-Wells/p/book/9780367497828
3	https://www.auctoresonline.org/journals/clinical-research-and-clinical-trials
4	https://www.who.int/health-topics/clinical-trials#tab=tab_1
5	https://www.cancerresearchuk.org/about-cancer/find-a-clinical-trial/what-clinical-trials-are/types-of-clinical-trials

Methods of Evaluation

Internal Evaluation	Continuous Internal Assessment Tests	25 Marks
	Assignments	
	Seminars	
	Attendance and Class Participation	
External Evaluation	End Semester Examination	75 Marks
	Total	100 Marks

Methods of Assessment

Recall (K1)	Simple definitions, MCQ, Recall steps, Concept definitions
Understand/Comprehend (K2)	MCQ, True/False, Short essays, Concept explanations, Short summary or overview
Application (K3)	Suggest idea/concept with examples, Suggest formulae, Solve problems, Observe, Explain.
Analyse (K4)	Problem-solving questions, Finish a procedure in many steps, Differentiate between various ideas, Map knowledge
Evaluate (K5)	Longer essay/ Evaluation essay, Critique or justify with pros and cons.
Create (K6)	Check knowledge in specific or offbeat situations, Discussion, Debating or Presentations.

Mapping with Programme Outcomes

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO 10	PO 11	PO 12	PO 13	PO 14
CO1	S	S	S		S									
CO2			S		S	S			S					
CO3		S		S		S			S					
CO4		S		S		S	S		S					
CO5				S				S	S		S		M	

Subject Code	Subject Name	Category	L	T	P	S	Credits	Inst. Hours	Marks		
									CIA	External	Total
22MB PGSE C1	Vermitechnology	Skill Enhancement Course 1	Y	-	-	-	2	2	25	75	100
Course Objectives											
CO1	Introduce the concepts of vermicomposting.										
CO2	Explain the physiology, anatomy and biology of earthworms.										
CO3	Acquire the knowledge of the vermicomposting process.										
CO4	Explain the trouble shooting, harvesting and packaging of vermin composts.										
CO5	Gain knowledge on applications of vermin composts and their value added products.										
UNIT	Details								No. of Hours	Course Objectives	
I	Introduction to Vermiculture - Definition, classification, history, economic importance- In sustainable agriculture, organic farming, earthworm activities, soil fertility & texture, soil aeration, water impercolation, decomposition & moisture, bait & food and their value in maintenance of soil structure. Its role in the bio transformation of the residues generated by human activity and production of organic fertilizers. Choosing the right worm. Useful species of earthworms. Local species of earthworms. Exotic species of earthworms. Factors affecting distribution of earthworms in soil.								6	CO1	
II	Earthworm Biology and Rearing - Key to identify the species of earthworms. Biology of <i>Eisenia fetida</i> . a) Taxonomy Anatomy, physiology and reproduction of Lumbricidae. b) Vital cycle of <i>Eisenia fetida</i> : alimentation, fecundity, annual reproducer potential and limiting factors (gases, diet, humidity, temperature, PH, light, and climatic factors). Biology of <i>Eudrilus eugeniae</i> . c) Taxonomy Anatomy, physiology and reproduction of Eudrilidae. d) Vital cycle of <i>Eudrilus eugeniae</i> : alimentation, fecundity, annual reproducer potential and limit factors (gases, diet, humidity, temperature, PH, light, and climatic factors).								6	CO2	
III	Vermicomposting Process - Feeds for Vermitech systems- Animal manures- Kitchen Waste and Urban waste- Paper pulp and card board solids- Compost and waste products- Industrial Wastes. Vermicomposting Basic process- Initial pre-composting phase- Mesophilic phase- Maturing and stabilization phase- Mechanism of Earthworm action. Methods of vermicomposting- a) windrows system; b) wedge system; c) container system-pits, tanks & cement rings; commercial model; beds or bins-top fed type, stacked type, d) Continuous flow system.								6	CO3	
IV	Vermicomposting - Trouble Shooting-Temperature-Aeration-Acidity- Pests and Diseases- Ants, rodents, Birds, Centipedes, sour crop, Mite pests. Odour problems. Separation techniques- Light Separation-Sideways Separation-Vertical Separation-								6	CO4	

	Gradual transfer. Harvesting Earthworms- manual method-migration method. Packing & Nutritional analysis of vermicompost.		
V	Applications of Vermiculture - Vermiculture Bio-technology, use of vermi castings in organic farming/horticulture, as feed/bait for capture/culture fisheries; forest regeneration. Application quantity of vermicompost in Agricultural fields- crops, fruits, vegetables & flowers. By-products and value-added products-Verm wash- vermicompost tea-vermi meal-enriched vermicompost-pelleted vermicompost.	6	CO5
	Total	30	

Course Outcomes

Course Outcomes	On completion of this course, students will;		
CO1	Compare and contrast the uses of vermicompost to the soil.		PO1, PO4, PO5, PO9,
CO2	Recommend different species of earthworms after acquiring knowledge on its biology.		PO1, PO4, PO6, PO9
CO3	Design the vermicomposting process.		PO1, PO4, PO6, PO7, PO8
CO4	Assess the Best Practices of Vermicomposting		PO6,PO7, PO8,PO9,
CO5	Recommend the applications of vermicompost to different soils and for different crops.		PO1, PO4, PO5,PO6, PO7

Text Books

1	Ismail S. A. (2005). The Earthworm Book, Second Revised Edition. Other India Press, Goa, India.
2	Rathoure A. K., Bharati P. K. and Ray J. (2020). Vermitechnology, Farm and Fertilizer. Vermitechnology, Farm and Fertilizer Discovery Publishing House Pvt Ltd.
3	Christy M. V. 2008. Vermitechnology, (1 st Edition), MJP Publishers.
4	The complete technology book on Vermiculture and Vermicompost with manufacturing Process, machinery equipment details and Plant Layout. AB Press.
5	Keshav Singh (2014). A Textbook of vermicompost: Vermiwash and Biopesticide.

References Books

1	Roy D. (2018). Handbook of Vermitechnology. Lambert Academic Publishing.
2	Kumar A. (2005). Verms and Vermitechnology, A.P.H. Publishing Corporation, New Delhi.
3	Lekshmy M. S., Santhi R. (2012). Vermitechnology, Sara Publications, New Delhi, India.
4	Edwards CA, Arancon NQ ShermanRL. (2011) Vermiculture Technology: Earthworms, Organic Wastes, and Environmental Management 1st edn.CRC Press.
5	Ismail, S.A. (1997). Vermicology-The Biology of Earthworm.1st edn. Orient longman.

Web Resources

1.	https://en.wikipedia.org/wiki/Vermicompost
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2.	http://stjosephs.edu.in/upload/papers/9567411a78c63d4ccfbbe85e6aa22840.pdf	
3.	https://www.kngac.ac.in/elearning-portal/ec/admin/contents/4_18K4ZEL02_2021012803204629.pdf	
4.	https://composting.ces.ncsu.edu/vermicomposting-2/	
5.	https://rodaleinstitute.org/science/articles/vermicomposting-for-beginners/	
Methods of Evaluation		
Internal Evaluation	Continuous Internal Assessment Tests	25 Marks
	Assignments	
	Seminars	
	Attendance and Class Participitation	
External Evaluation	End Semester Examination	75 Marks
	Total	100 Marks
Methods of Assessment		
Recall (K1)	Simple definitions, MCQ, Recall steps, Concept definitions	
Understand / Comprehend (K2)	MCQ, True/False, Short essays, Concept explanations, Short summary or overview	
Application (K3)	Suggest idea/concept with examples, Suggest formulae, Solve problems, Observe, Explain	
Analyse (K4)	Problem-solving questions, Finish a procedure in many steps, Differentiate between various ideas, Map knowledge	
Evaluate (K5)	Longer essay/ Evaluation essay, Critique or justify with pros and cons	
Create (K6)	Check knowledge in specific or offbeat situations, Discussion, Debating or Presentations	

Mapping with Programme Outcomes

	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO10	PO11	PO12	PO13	PO14
CO1	S			M	S				S					
CO2	S			M		S			S					
CO3	S			S		S	S	S						
CO4						S	S	S	S					
CO5	S			M	S	M	S							

Subject Code	Subject Name	Category	L	T	P	S	Credits	Inst. Hours	Marks		
									CIA	External	Total
22M BPG CT7	Soil and Environmental Microbiology	Core Course VII	Y	Y	-	-	4	6	25	75	100
Course Objectives											
CO1	Explain the role of microorganisms in soil fertility.										
CO2	Discuss the benefits of interactions among soil microbes and acquire awareness about microbes as biofertilizers and biocontrol agents.										
CO3	Create awareness. about components of environment, environmental pollution, and detection methods.										
CO4	Acquire in depth knowledge about solid and liquid waste treatments.										
CO5	Develop knowledge about organic matter degradation, bioremediation, and the environment risk assessment.										
UNIT	Details								No. of Hours	Course Objectives	
I	Soil Microbiology– Soil as Microbial Habitat, Soil profile and properties, Soil formation, Diversity, and distribution of major group of microorganisms in soil. Quantification of soil microflora, role of microorganism in soil fertility. Mineralization of Organic & Inorganic Matter in Soil. Biological Nitrogen fixation- Chemistry and Genetics of BNF.								20	CO1	
II	Microbial Interactions - Mutualism, Commensalism, Amensalism, Synergism, Competition, Rhizosphere- Rhizosphere effect, Mycorrhizae – Types, Endophytes, PGPR- Plant growth promoting bacteria– symbiotic (<i>Bradyrhizobium</i> , <i>Rhizobium</i> , <i>Frankia</i>), Non-Symbiotic (<i>Azospirillum</i> , <i>Azotobacter</i> , Mycorrhizae, MHBs, Phosphate solubilizers, algae), Novel combination of microbes as biofertilizers, PGPRs.								20	CO2	
III	Components of Environment: Hydrosphere, lithosphere, atmosphere, and biosphere – definitions with examples; Energy								15	CO3	

	flow in the ecosystem- Carbon, Nitrogen, Sulfur and Phosphorous cycles. Physical factors affecting distribution of microorganisms in various environments. Predisposing factors for Environmental diseases – infectious (water and air borne) and pollution related, spread and control of these diseases. Treatment and safety of drinking (potable) water, methods to detect potability of water samples. Space microbiology - Microbiological research in space environment.		
IV	Waste management – Solid waste - Types - management - Factors affecting solid waste generation rates. Industrial effluent treatment, primary, secondary, tertiary, and advanced treatment process. Quality assessment of decontaminated matters and other biological effluents. Biological reference standards. Utilization of Solid Waste as Food, Feed and Fuel- Composting, Vermicomposting, Bio manure and Biogas production. E waste management.	15	CO4
V	Degradation of organic matter - lignin, cellulose, hemicellulose, pectin, common pesticides- herbicides (2,4-D) and pesticides (DDT), heavy metals. Biodegradation of Xenobiotics - Recalcitrant Halocarbons, Recalcitrant TNTs, PCBs and Synthetic polymers. Biodegradation of Hydrocarbons. Biodeterioration of Textiles and Leather. Pollution Control Bodies and Environmental laws in India. Environmental impact assessment, EIA guidelines, US Environment protection Agency norms.	20	CO5
	Total	90	

Course Outcomes

Course Outcomes	On completion of this course, students will;	
CO1	Depict diversity and significance of soil microbes and predict the role of microbes in biological nitrogen fixation.	PO1
CO2	Utilize the knowledge of microbial interactions, with beneficial application of biofertilizers for sustainable agriculture and benefits of biopesticides.	PO1, PO7, PO8
CO3	Explain the different types of microorganisms in water. Identify the causes of water pollution and the methods for quality assessment of water and control of water borne diseases.	PO1, PO5, PO6, PO7, PO8
CO4	Apply knowledge about waste treatments and microbial decomposition and bio-remediation process in environmental cleanup.	PO1, PO5
CO5	Plan a clear approach on environmental issues. Control pollution and explain protection laws to public.	PO1, PO5

Text Books

1.	Subba Rao. N. S. (2017). Soil Microbiology. (5 th Edition). MedTech Publishers.
2.	Daniel. C. J. (2006). Environmental Aspects of Microbiology. (2 nd Edition). Bright Sun Publications.
3.	Rangaswami. G. and Mahadevan. A. (2006). Diseases of Crop Plants in India. (4 th Edition). Prentice–Hall of India Pvt. Ltd.

4.	Sharma P. D. (2010). Microbiology and Plant pathology. (2 nd Edition). Rastogi Publications.	
5.	Subba Rao. N.S. (2005). Soil microorganisms and Plant Growth. (4 th Edition). Oxford and IBH Publishing Pvt. Ltd.	
References Books		
1.	Pepper I. L., Gerba C. P. and Gentry T. J. (2014). Environmental Microbiology (1 st Edition). Academic Press, Elsevier.	
2.	Bitton, G. (2011). Wastewater Microbiology. (4 th Edition). Wiley-Blackwell.	
3.	Bridgewater L. (2012). Standard Methods for the Examination of Water and Wastewater. American Public Health Association.	
4.	Shrivastava A.K. (2003). Environment Auditing. A. P. H. Publishing Corporation.	
5.	Tinsley, S. and Pillai, I. (2012). Environmental Management Systems – Understanding Organizational Drivers and Barriers. Earthscan.	
Web Resources		
1.	https://academic.oup.com/femsec/article/93/5/fix044/3098413	
2.	http://www.fao.org/3/t0551e/t0551e05.htm	
3.	www.environmentshumail.blogspot.in/	
4.	https://www.frontiersin.org/articles/10.3389/fpls.2017.01617/full	
5.	https://serc.carleton.edu/microbelife/index.html	
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